Author for correspondence:
Changbong Hyeon
e-mail: hyeoncb@kias.re.kr

Glycolytic oscillations under periodic drivings

Pureun Kim and Changbong Hyeon

Korea Institute for Advanced Study, Seoul 02455, Korea

CH, 0000-0002-4844-7237

In many living organisms displaying circadian rhythms, the intake of energy often occurs in a periodic manner. Glycolysis is a prototypical biochemical reaction that exhibits a self-sustained oscillation under continuous injection of glucose. Here we study the effect of periodic injection of glucose on the glycolytic oscillation from a dynamical systems perspective. In particular, we employ Goldbeter's allosteric model of phosphofructokinase as a model system for glycolytic oscillations, and explore the effect of periodic substrate influx of varying frequencies and amplitudes by building the phase diagrams of Lyapunov exponents and oscillatory periods. When the frequency of driving is tuned around the harmonic and sub/super-harmonic conditions of the natural frequency, the system is entrained to a frequency-locked state, forming an entrainment band that broadens with an increasing amplitude of driving. On the other hand, if the amplitude is substantial, the system may transition, albeit infrequent, to a chaotic state which defies prediction of dynamical behaviour. Our study offers in-depth understandings into the controllability of glycolytic oscillation as well as explaining physical underpinnings that enable the synchronous oscillations among a dense population of cells.

1. Introduction

Glycolysis is one of the metabolic processes essential for all living organisms. It is an anaerobic process that catalyses the transformation of one glucose molecule into two pyruvates, each resulting in a net production of two adenosine triphosphates (ATPs) and a reduction of nicotinamide adenine dinucleotide (NAD⁺) to NADH, which are later used in the processes of Kreb cycle and electron transfer cycle to generate more ATPs under aerobic condition [1]. Notably, in yeast and heart muscle cell extracts [2–4] and in pancreatic β-cells [5–7], the products of glycolysis exhibit sustained oscillations even when the influx of glucose to the system is constantly maintained.

Among various models and mechanisms developed for describing glycolytic oscillations [8–12], the positively regulated autocatalytic–allosteric action of phosphofructokinase (PFK) has been suggested as a core mechanism for the generation of the sustained oscillation [4,8,13,14]. Specifically, PFK is a phosphotransferase that catalyses ATP hydrolysis, so as to convert fructose 6-phosphate (F6P) into fructose 1,6-biphosphate (FBP). Binding of one of the reaction products (adenosine diphosphate (ADP)) to the regulatory sites of PFK positively regulates the allosteric transition of PFK into a catalytically more competent state, which generates oscillations in substrate and product concentrations. In experiments using yeast cell extracts, sustained oscillations of NADH concentration were observed as long as the influx of glucose and the degradation of product were maintained in a proper range [4,15].

From the perspective of dynamical systems of continuous variables in two dimensions, the sustained oscillation of substrate and product concentrations ($[S]$ and $[P]$) can be understood as a stable limit cycle formed around an unstable fixed point, which is ensured by the Poincaré–Bendixon theorem [16]. An introduction of third variable that changes over time, such as a non-autonomous time-varying influx of glucose to the system, however,

substantially increases the complexity of the resulting dynamics. In particular, the external periodic driving effectively puts dynamical trajectories on the phase space surface of a torus [17,18]. Boiteux *et al.* [4] carried out an experimental study to explore the effect of stochastic and *periodic* injection of glucose on the glycolytic oscillation, and observed subharmonic entrainments of the system for the case of periodic injection (see appendix for the effect of stochastic forcing). That is, when the period of external driving (T_{ext}) relative to the autonomous period of the system (T_o) was over a certain range, the system was entrained towards a new period of $T_{obs} = nT_{ext}$ ($n = 1$, 2, 3, …). Meanwhile, if the system was not entrained, the external driving gave rise to an irregular trajectory.

Our study on the PFK model under periodic influx of substrate can be considered as a theoretical extension of Boiteux *et al.*'s experimental work [4]. We find that the perturbation leads to extremely rich dynamical behaviours. By varying the frequency ($\omega_{ext} = 2\pi/T_{ext}$) and amplitudes ($\varepsilon$) of the external driving, we calculate two-dimensional phase diagrams of (i) the Lyapunov exponent (λ) and (ii) the resulting oscillation period (T_{obs}), which not only clarify the condition that leads to entrainment, but also help differentiate between chaotic and quasi-periodicity orbits. Our study offers in-depth understandings into the controllability of glycolytic oscillation by illuminating its dynamical responses and modulations under periodic drivings.

2. Phosphofructokinase model of glycolytic oscillation

The PFK is an oligomeric enzyme that retains a catalytic and a regulatory site in each of n subunits, where the value of n ($= 2$, 4, 8) varies with the species. At low substrate concentration, the PFK is predominantly in the tense (T) state, but high concentration of substrate shifts the pre-equilibrated population of enzyme in T state to the *catalytically competent* relaxed (R) state. The PFK in the R state is capable of converting F6P into FBP via phosphorylation while catalysing the ATP hydrolysis. A reaction product, ADP, cooperatively binds to the n ($= 8$ for yeast) distinct regulatory sites available in the R state and positively regulates the $T \rightarrow R$ transition of PFK. Thus, with an increasing substrate concentration, PFK has more chances to exhibit its catalytic power, which accelerates with the amount of ADP until the n regulatory sites are half filled by ADP, but saturates when all the sites are filled up, giving rise to an activity curve with a sigmoidal shape.

The autocatalytic–allosteric model of glycolysis processed by PFK is formulated using a system of ordinary differential equations (ODEs) [8,19],

$$\left.\begin{aligned} \frac{d[S]}{dt} &= \nu_0 - k \sum_{i=0}^{n} \sum_{j=0}^{n} i[R_{ij}] \\ \text{and} \quad \frac{d[P]}{dt} &= k \sum_{i=0}^{n} \sum_{j=0}^{n} i[R_{ij}] - k_d[P]. \end{aligned}\right\} \quad (2.1)$$

The ODEs describe the mass balance of the substrate and product concentrations. The substrate S supplied to the system with a rate ν_0 is converted by PFK to a product P with a rate proportional to the occupation number of the substrate in the catalytic sites of R state, $k \sum_{i=0}^{n} \sum_{j=0}^{n} i[R_{ij}]$ where k denotes

the conversion rate and R_{ij} denotes a microstate of R state in which i ($= 0, 1, …, n$) catalytic sites and j ($= 0, 1, …, n$) regulatory sites are occupied by the substrate and product, respectively. The product P undergoes uni-molecular degradation with the rate k_d from the system. In writing the ODEs in equation (2.1), it is assumed that the conformational changes of PFK enzyme into its subpopulations of T or R states occur in much faster time scales than the time scale associated with the variations in substrate and product concentration. As a consequence, the corresponding concentrations of the subpopulations $[T_i]$ and $[R_{ij}]$ are pre-equilibrated and can be expressed as a function of the substrate and product concentration ($[S]$ and $[P]$) (see equations (2.2) and (2.3)). The PFK model, proposed by Goldbeter & Lefever [8], is built based on the Monod–Wyman–Changeux (MWC) model [20–22], which is the workhorse of protein allostery [22–25], along with a set of key parameters:

— $L \equiv [T_0]/[R_0]$, where $R_0 \equiv R_{00}$ is the ratio of T and R states in the absence of substrate.
— $K_T^S (= k_{T,off}^S / k_{T,on}^S)$ and $K_R^S (= k_{R,off}^S / k_{R,on}^S)$ are the dissociation constants or the binding affinities of substrate (S) to the T and R states, respectively.
— $c \equiv K_R^S / K_T^S$, the ratio of binding affinities in R and T states, specifies the preference of substrate binding to the R against the T state.

In the PFK model, the concentration of PFK subpopulation in the T state bound by i substrates ($0 \leq i \leq n$) can be expressed as

$$[T_i] = L \binom{n}{i} \left(\frac{c[S]}{K_R^S}\right)^i \times [R_0], \quad (2.2)$$

and similarly the concentration of PFK subpopulation in the R state bound by i substrates in the catalytic sites and j product molecules in the regulatory sites is given as

$$[R_{ij}] = \binom{n}{i} [\alpha([S])]^i \binom{n}{j} [\gamma([P])]^j \times [R_0], \quad (2.3)$$

where $\alpha([S]) \equiv [S]/(K_R^S + k/k_{R,on}^S)$ and $\gamma([P]) \equiv [P]/K_R^P$ with K_R^P denoting the binding affinity of product (ADP) to the regulatory site of the R state. Using equations (2.2), (2.3) and the total enzyme concentration $E_{tot} = \sum_i [T_i] + \sum_{i,j} [R_{ij}] = \{L(1 + c[S]/K_R^S)^n + (1 + \alpha([S]))^n (1 + \gamma([P]))^n\}[R_0]$, one can express $\sum_{i=0}^{n} \sum_{j=0}^{n} i[R_{ij}]$ in terms of $[S]$ and $[P]$ as

$$\sum_{i=0}^{n} \sum_{j=0}^{n} i[R_{ij}] = \frac{n\alpha([S])(1 + \alpha([S]))^{n-1}(1 + \gamma([P]))^n E_{tot}}{L(1 + c[S]/K_R^S)^n + (1 + \alpha([S]))^n (1 + \gamma([P]))^n}, \quad (2.4)$$

which clarifies that equation (2.1) is a set of ODEs with two variables, $\dot{\mathbf{x}} = \mathbf{F}(\mathbf{x})$ with $\mathbf{x} = ([S], [P])$.

In our earlier work [26], we used the parameters, $k_{R,on}^S = 2000 /\text{mM s}$, $K_R^S = 0.05 \text{ mM}$, $K_R^P = 0.025 \text{ mM}$ [27–30] and $c = K_R^S / K_T^S = 0.01$ which ensures preferential binding of substrate to the R state. Since the ATP hydrolysis time due to ATPase activity is typically $\gtrsim \mathcal{O}(1)$ msec [31], we used $k = 500 \text{ s}^{-1}$ for the rate of catalysis. In addition, by specifically considering the yeast PFK that adopts an octameric form ($n = 8$), we use the allosteric constant $L = 4 \times 10^9$ which is 3 orders of magnitude greater than the value known for the tetramer ($n = 4$) [29]. In [26], we studied the phase diagrams of various quantities as a function of constant influx of substrate ν_0 and the degradation rate k_d, identifying the parameter space
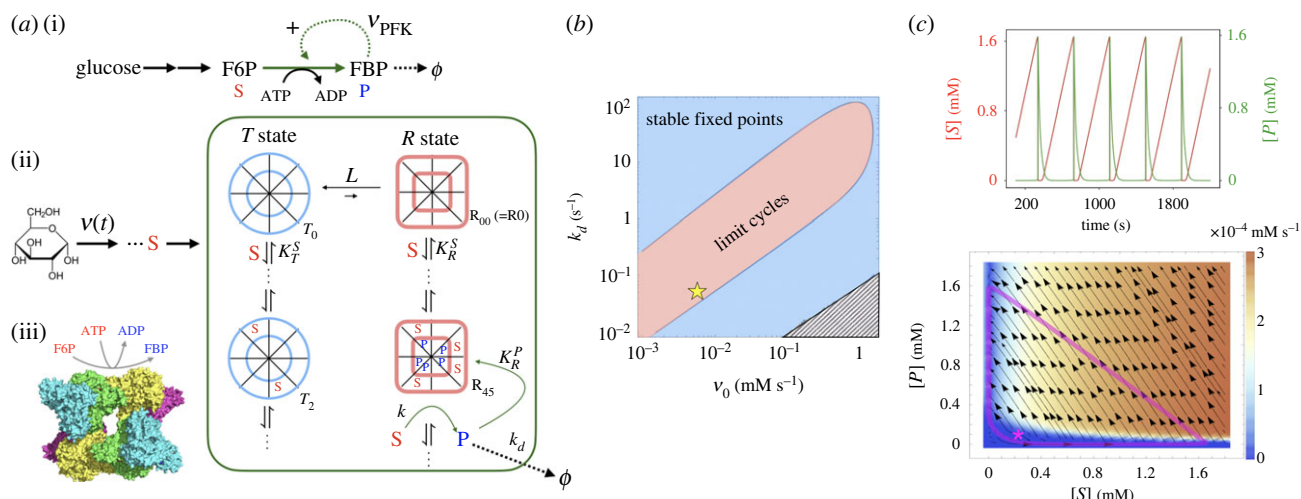
Figure 1. Allosteric model for phosphofructokinase. (a) (i) The schematic of glycolytic pathway, highlighting the F6P → FBP conversion regulated by PFK that undergoes positive allosteric activation. (ii) The more detailed schematic highlighting the allosterism of PFK catalysis. Each subunit of PFK has catalytic and regulatory sites, where substrate (S = F6P) and product (P = ADP) bind, respectively. The oscillatory dynamics of PFK model is realized in a setting of open thermodynamic system, where glucose molecules, which lead to the production of F6P, are injected at the rate $v(t)$ and the product (P) degrades at the rate of k_d. At low substrate concentration ($[S] < K_T^S$), the enzyme is mainly in the T state. But, with increasing substrate concentration, conformational transition to the R state occurs, increasing the chance of enzymatic activity. (iii) The octameric structure ($n = 8$) of PFK (left bottom corner), which catalyses the conversion of F6P to FBP via the ATP hydrolysis. (b) Phase diagram of dynamical behaviour of PFK model as a function of constant injection rate $v(t) = v_0$ and degradation rate k_d. A set of parameter values relevant for octameric PFK of yeast ($L = 4 \times 10^9$, $k = 500$ s^{-1}, $v_0 = 0.005$ mM s^{-1}, $k_d = 0.05$ s^{-1}, $c = 0.01$, $E_{tot} = 5 \times 10^{-4}$ mM and $n = 8$), which corresponds to the location marked with a yellow star on the phase diagram, yields a stable limit cycle with a period of $T_o = 388.6$ s. (c) Anti-phasic oscillations of $[S](t)$ and $[P](t)$ (top), and the phase portrait (bottom) with the velocity fields $\boldsymbol{v}([S], [P]) = (\mathrm{d}[S]/\mathrm{d}t, \mathrm{d}[P]/\mathrm{d}t)$ along with the steady state trajectory (thick magenta line) around the unstable fixed point ($[S^*], [P^*]) = (0.22$ mM, 0.10 mM) marked by an asterisk. The colour-code in the phase portrait depicts the size of the velocity field, $\|\boldsymbol{v}([S], [P])\|$.

associated with the self-sustained oscillations (figure 1b). For the parameter values relevant for octameric PFK of yeast ($L = 4 \times 10^9$, $k = 500$ s^{-1}, $v_0 = 0.005$ mM s^{-1}, $k_d = 0.05$ s^{-1}, $c = 0.01$, $E_{tot} = 5 \times 10^{-4}$ mM and $n = 8$), which yields a stable oscillation with a period of $T_o = 388.6$ s, we find the system poised at a thermodynamically optimal point with minimal entropy production rate [26,32] in the limit cycle region near the phase boundary (the point marked with the yellow star in figure 1b), corresponding to the state above the Hopf bifurcation point [26].

Here, we explore how the oscillatory behaviour of the PFK model is modulated under a periodic influx of substrate $v(t)$ satisfying

$$v(t) = \begin{cases} v_0 + \varepsilon \sin(\omega_{ext}t), & \text{for } v(t) \geq 0 \\ 0, & \text{for } v(t) < 0 \end{cases} \quad (2.5)$$

with $\omega_{ext} = 2\pi / T_{ext}$. For $\varepsilon > v_0$, $v(t)$ is set to 0 if $v(t) < 0$, which renders $v(t)$ a strong periodic pulse to the system.

The main set of ODEs, equation (2.1) with equation (2.4), was numerically integrated to generate time trajectories of $[S](t)$ and $[P](t)$ using the backward differentiation formula (BDF) along with the time-periodic drivings specified in equation (2.5).

3. Results

## 3.1. Phase diagram, entrainment and quasi-periodic orbits

Our calculation of the two-dimensional diagram for the PFK model under external periodic driving is summarized in figures 2 and 3. Figure 2a depicts the Lyapunov exponent evaluated for dynamic time trace as a function of the parameters, Ω and ε that vary in the ranges of $0 \leq \Omega \leq 5$ and

$0 \leq \varepsilon \leq 1.2v_0$ (see Methods for the detail of evaluating Lyapunov exponent). Here, the Ω is the ratio of the system's natural frequency of oscillation to the frequency of external driving, i.e. $\Omega = \omega_o/\omega_{ext} = T_{ext}/T_o$. In figure 2b, the period of entrained dynamics T_{obs} is calculated on each grid point, if T_{obs} is resolved within our simulation time (i.e. $T_{obs} < T_s = 10^4 \times T_{ext}$); and no specific value of T_{obs} is assigned to the grid point if the dynamics is either quasi-periodic or chaotic (see equation (5.16) in Methods). Some of the time traces at their stationary state are plotted in figure 2c. The trajectories, marked with star symbols in the diagram of λ in figure 2a and $[S](t)$ and $[P](t)$ in figure 2c, are:

— (red) frequency-locked orbit entrained to $T_{obs} \approx 389$ s generated with $\varepsilon = 0.6v_0$ and $\Omega \approx 1$ ($T_{ext} = T_o \approx 389$ s);
— (green) frequency-locked orbit entrained to $T_{obs} \approx 389$ s generated with $\varepsilon = 0.4v_0$ and $\Omega \approx 1/3$ ($T_{ext} \approx 129$ s, $T_o \approx 389$ s);
— (cyan) *effectively* quasi-periodic orbit generated with $\varepsilon = 0.3v_0$ and $\Omega = 2.797$; and
— (yellow) chaotic orbit generated with $\varepsilon = 0.92v_0$ and $\Omega = 2.393$.

In the limit of small perturbation ($\varepsilon \to 0$), if the parameter Ω is rational (T_{ext} is commensurate with T_o), the system is entrained into a frequency-locked state with a period T_{obs}, which corresponds to the least common multiple (LCM) of T_{ext} and T_o, i.e. $T_{obs} = \text{LCM}(T_{ext}, T_o)$. By contrast, if the ratio of frequencies is irrational (T_{ext} is incommensurate with T_o), the resulting dynamics is quasi-periodic to display an orbit with an indefinite period ($T_{obs} \to \infty$). In practice, if $T_{obs} = \text{LCM}(T_{ext}, T_o)$ acquired from a rational Ω were much greater
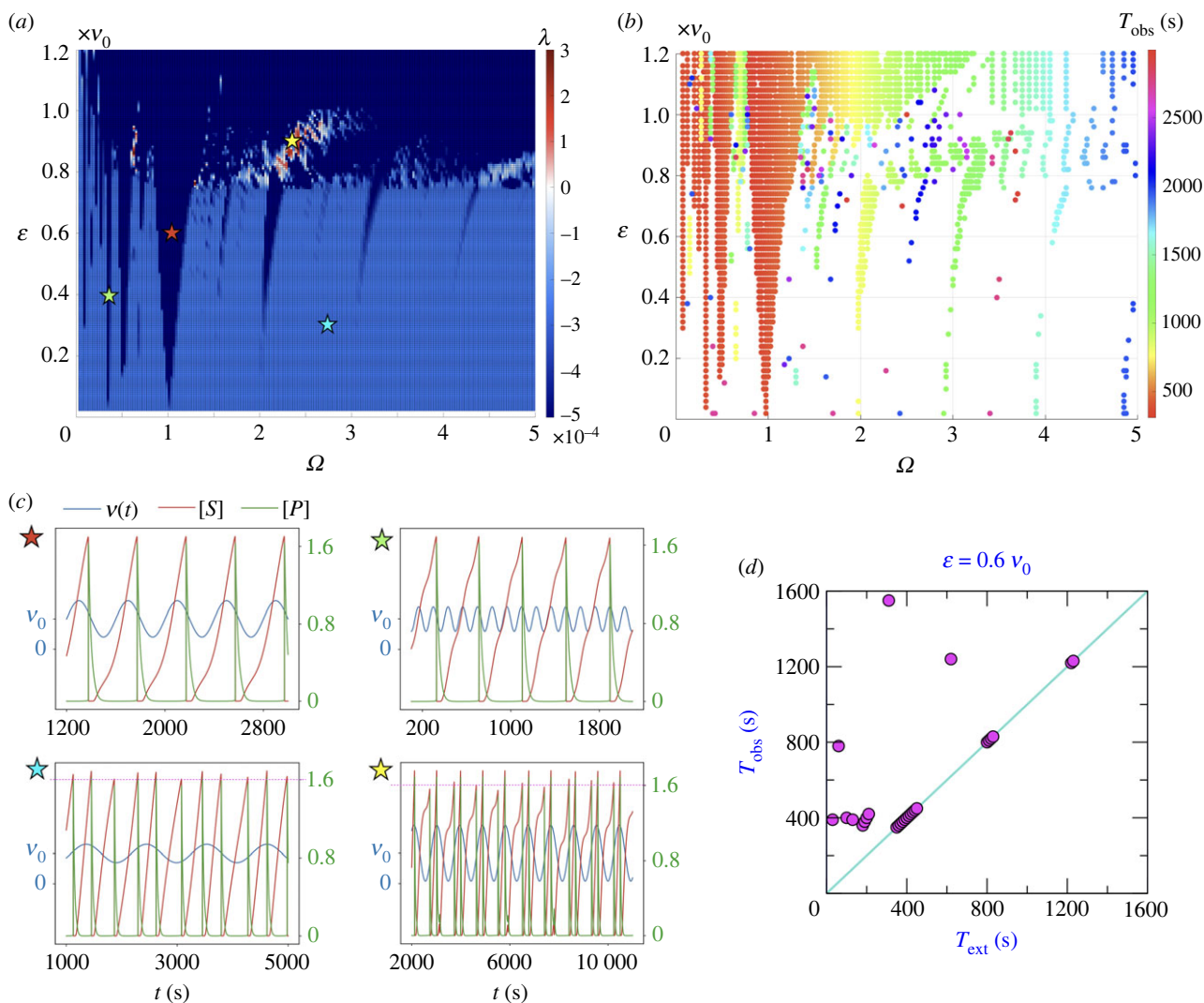
Figure 2. The two-dimensional diagrams of (*a*) the Lyapunov exponent λ and (*b*) the observed oscillation period T_{obs} within the entrainment bands as a function of ε and Ω. The oscillation periods less than 3000 s, which imply that systems are entrained into a frequency-locked state, are depicted with filled circles in rainbow colour. (*c*) The actual time trajectories at parameter values marked with star symbols in different colours. Substrate concentration $[S](t)$ in red and product concentration $[P](t)$ in green in response to periodic driving $\nu(t)$ in blue. The dotted lines in magenta on the panels marked with cyan and yellow stars are drawn to highlight the irregularity of spikes in each interval of the periodic driving ($\nu(t)$). The trajectories generated at different sets of parameters, ε and Ω, marked with star symbols, are: (red) frequency-locked state entrained to $T_{obs} \approx 400$ s ($\varepsilon = 0.6\nu_0$, $\Omega \approx 1$); (green) frequency-locked state entrained to $T_{obs} \approx 400$ s ($\varepsilon = 0.4\nu_0$, $\Omega \approx 1/3$); (cyan) *effectively* quasi-periodic state ($\varepsilon = 0.3\nu_0$, $\Omega = 2.797$); (yellow) chaotic state ($\varepsilon = 0.92\nu_0$ and $\Omega = 2.391$). (*d*) The entrainment period (T_{obs}) as a function of the period of external driving (T_{ext}) calculated at $\varepsilon = 0.6\nu_0$. The clusters of points at $T_{obs} \approx 400$ s correspond to the periods of entrainment calculated in the Arnold tongues of subharmonic ($T_{ext} \approx T_o/2$, $T_o/3$, $T_o/4$) and harmonic ($T_{ext} \approx T_o$) entrainments. The clusters of points at $T_{obs} \approx 800$ s and $T_{obs} \approx 1200$ s are due to the Arnold tongues formed under the super-harmonic entrainments. The cyan line corresponds to the condition of $T_{obs} = T_{ext}$.

such that $T_{obs} \gg T_{ext}$, the corresponding frequency-locked state would display an oscillatory orbit with a long period. The corresponding trajectory densely fills the state space and is effectively indistinguishable from the dynamics of a truly quasi-periodic state generated from an irrational Ω. In such a case, we regard the trajectory *effectively quasi-periodic*.

Difference between periodic and (effectively) quasi-periodic trajectories is more clearly grasped by plotting dynamic time traces on a polar coordinate and their piercings on a surface of the Poincaré section (figure 3). Dynamic trajectories of frequency-locked states with $\Omega \approx 1$ ($T_{ext} \approx 389$ s) and $\Omega \approx 1/3$ ($T_{ext} \approx 129$ s), plotted as a function of phase variable $\theta(t) = \omega_{ext}t \mod 2\pi$ with $\omega_{ext} = 2\pi/T_{ext}$ on the polar coordinate, form closed loops with a single (figure 3*a*, left) and three revolutions (figure 3*b*, left), displaying a single (figure 3*a*, right) and three piercings (figure 3*b*, right) on the Poincaré section at $\theta(t_n) = 0$, respectively. In comparison, the trajectory

of an effectively quasi-periodic state with $\Omega = 2.797$ ($T_{ext} = 1085$ s, the trajectory marked with cyan star in figures 2 and 3), which results in an orbit with $T_{ext} \approx 420\,980$ s, almost fully covers the phase space on the polar coordinate as well as giving rise to the continuous piercings on the corresponding Poincaré section.

A finite amplitude ($\varepsilon \neq 0$) coupled to the oscillatory mode of the system expands the parameter range of Ω for the frequency-locked state, and thus widens the entrainment bands, forming the Arnold tongues [33–36] (figure 2*b*). Consistent with the experiment by Boiteux *et al.* [4], the subharmonic entrainments to $T_{obs} = nT_{ext} \approx T_o \approx 389$ s for $\Omega (= T_{ext}/T_o) \sim 1/n$ with $n = 2$, 3, 4, ... are confirmed for $\Omega \lesssim 1$ (figure 2*b*). For $\Omega > 1$, we observe super-harmonic entrainments to $T_{obs} \approx 1200$ s at $\Omega \approx 3/2$, $T_{obs} \approx 800$ s at $\Omega \approx 2$, $T_{obs} \approx 1200$ s at $\Omega \approx 3$, and $T_{obs} \approx 1600$ s at $\Omega \approx 4$ (figure 2*b*). The width of the Arnold tongue is the greatest for the harmonic entrainment, i.e. $\Omega \sim 1$.
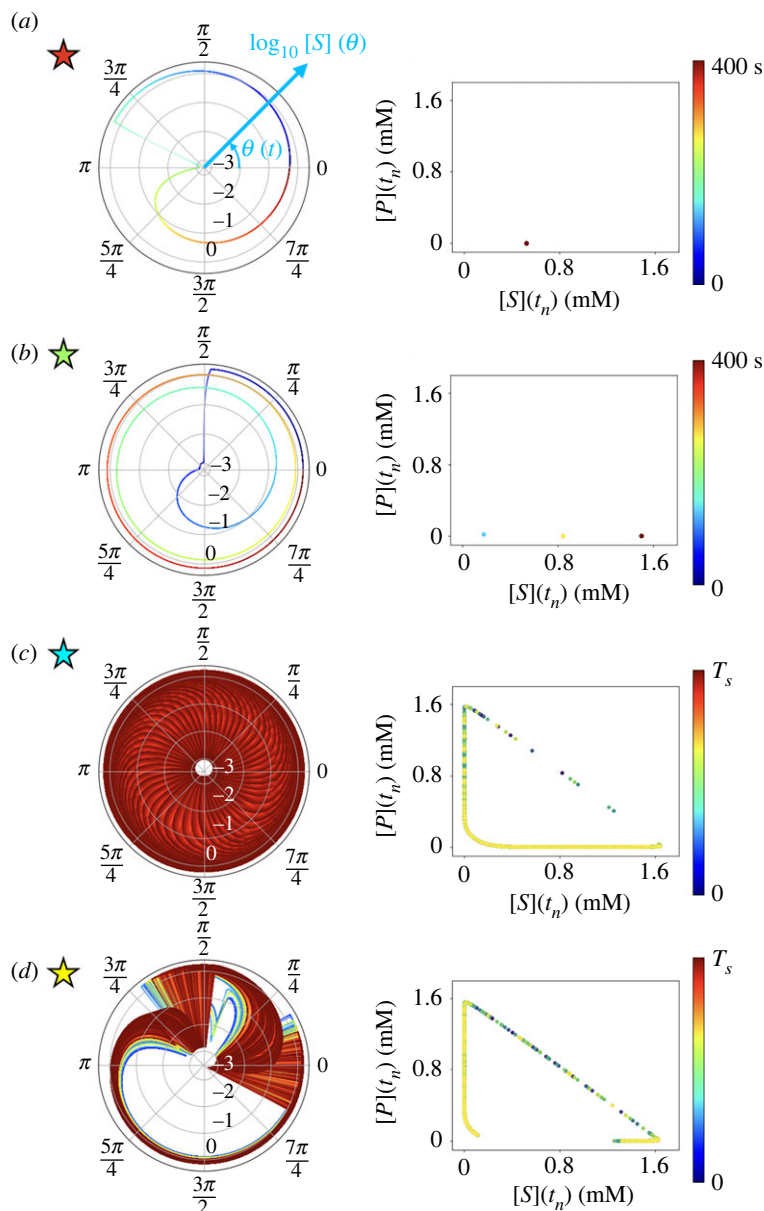
Figure 3. Analysis of the trajectories at different parameter values, marked with star symbols in red (a), green (b), cyan (c), and yellow (d) in figure 2a,c. Steady state trajectory of $[S](\theta)$ (radial direction in logarithmic scale) as a function of phase angle $\theta = \omega_{ext}t \bmod 2\pi$ depicted on the polar coordinate (left), the intersections of trajectory across the Poincaré section ($[S](t_n)$, $[P](t_n)$) at t_n satisfying $\theta(t_n) \bmod 2\pi = 0$ (right). The trajectories and the intersections across the Poincaré section are colour-coded in time over $0 \le t \le T_{obs}$ for frequency-locked states and for simulation time ($0 \le t \le T_s$ for quasi-periodic and chaotic states.

In principle, unless $\lambda > 0$, all the time traces generated at each grid point of rational Ω (figure 2) are still characterized with a finite periodicity $T_{obs} < \infty$. However, outside the entrainment bands, the two-dimensional phase diagram is in fact densely populated by quasi-periodic states since the density of irrational Ω is significantly higher than that of rational Ω. The Lebesgue measure of irrational Ω, giving rise to quasi-periodic orbits, is 1 for $\varepsilon \to 0$ [17,18]. Meanwhile, an increasing value of ε leads to expansions of the entrainment bands, which in turn reduces the region associated with the quasi-periodicity. Figure 2d, which visualizes the periods of entrainment (T_{obs}) for a given period of external driving (T_{ext}) at $\varepsilon = 0.6\nu_0$, hints at the winding number [34,35] of the map associated with our model.

3.2. Chaotic state

The conditions that give rise to a chaotic orbit are of great interest. The trajectories of $[S](t)$ and $[P](t)$ at a chaotic state

($\varepsilon = 0.92\nu_0$, $\Omega = 2.391$) which display a train of spikes with irregular height (figure 2c, yellow star) are similar to those with quasi-periodicity (figure 2c, cyan star), and the patterns of piercings on the Poincaré section are also qualitatively similar (figure 3c,d). However, together with the sign of Lyapunov exponent, chaotic orbit is differentiated from quasi-periodic orbit when the trajectory is drawn on the polar coordinate of phase angle. The phase space represented in the polar coordinate (figure 3) is only partly filled with the chaotic orbit, whereas it is fully filled with the quasi-periodic orbit.

The chaotic states with positive Lyapunov exponents ($\lambda > 0$) are sporadically identified in the phase diagram. In figure 2a, they are localized in the vicinity of $\Omega \approx 0.5$ and $\Omega \approx (2-2.5)$ for $\varepsilon \approx (0.8-1)\nu_0$. Figure 4a extends the phase diagram displayed in figure 2a to a wider range of ε. It is clear from figure 4a that while chaotic states are scarce under strong driving ($\varepsilon \gtrsim \nu_0$), they are still found at $\varepsilon \approx 4\nu_0$ and $7\nu_0$ in the range of $0 < \Omega \lesssim 0.3$. The inset of figure 4a magnifies the region of phase diagram where chaotic states are frequently observed,
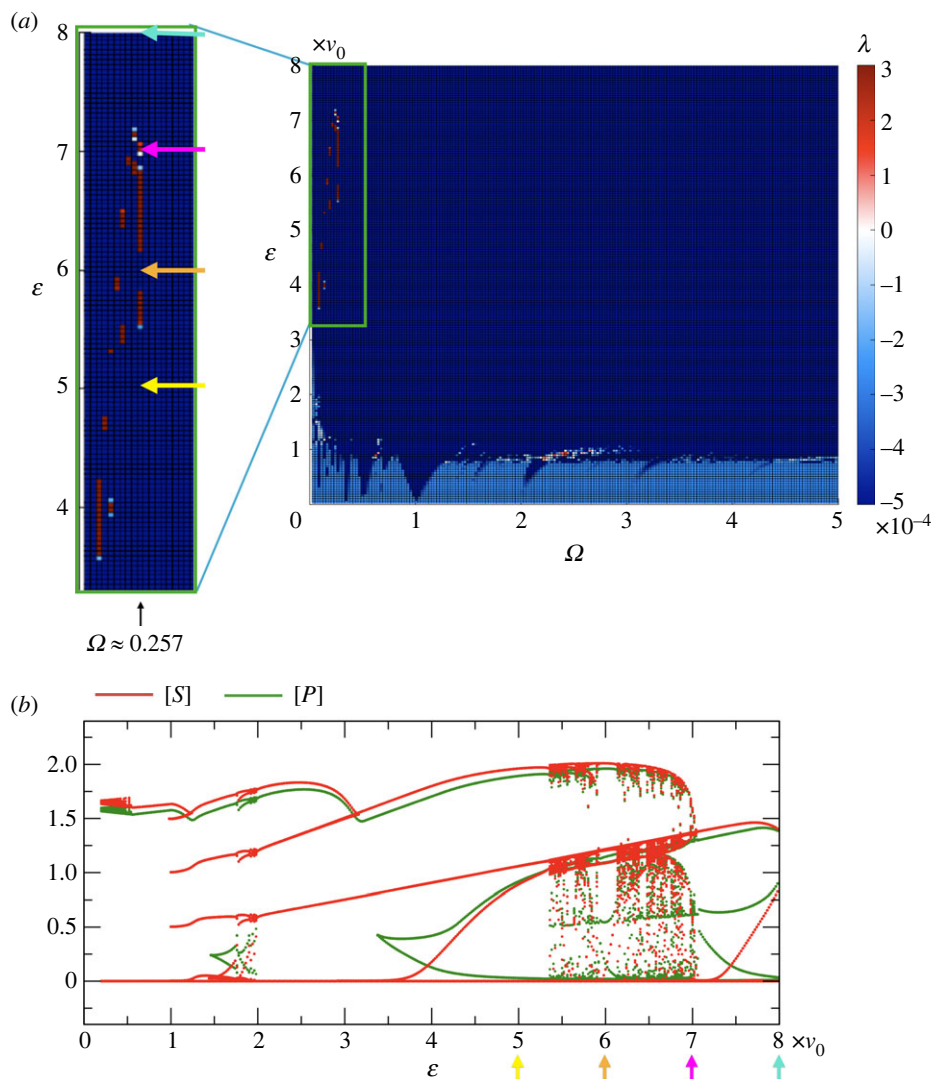
Figure 4. Transition to chaos demonstrated for ε extended to large values at fixed Ω $(= T_{ext}/T_o = 100/389) \approx 0.257$. (a) Phase diagram of the Lyapunov exponent λ extended to large ε. The inset on the left enlarges the region of phase diagram with chaotic states. The trajectories of $[S](t)$ and $[P](t)$ at four distinct sets of parameter values, indicated by the arrows, are demonstrated in figure 5. (b) Chaotic maps for $[S](t)$ and $[P](t)$ as a function of ε calculated at fixed $\Omega \approx 0.257$.

and the chaotic maps at fixed $\Omega = 0.257$ (figure 4b) visualize qualitative changes of the periodic orbits, $[S](t)$ and $[P](t)$ with increasing ε. The periodic orbits generated at four specific values of ε marked with the arrows in different colours in the inset of figure 4a,b are shown in figure 5. Straightforwardly gleaned from the number of piercings in the Poincaré section (the panels in the rightmost column of figure 5), the system is in the frequency-locked states of period-3 ($\varepsilon = 5\nu_0$), period-5 ($\varepsilon = 6\nu_0$) and period-2 cycles ($\varepsilon = 8\nu_0$); however, it displays a chaotic behaviour at $\varepsilon = 7\nu_0$. The irregularities in the size and the number of spikes of the chaotic orbit ($\varepsilon = 0.7\nu_0$) are seen in the leftmost panel in figure 5, and the other remaining panels of figure 5 capture the characteristic features of chaotic states.

3.3. Mapping onto the circle map
To analyse the model of glycolytic oscillation under the periodic driving more systematically, we recast the original set of ODEs in the following form:

$$\left.\begin{array}{l} \partial_t[S] = F_1([S],[P]) + \varepsilon u(t) \\ \partial_t[P] = F_2([S],[P]), \end{array}\right\} \quad (3.1)$$

where $u(t) = \sin(\omega_{ext}t) = u(t + T_{ext})$ is a T_{ext}-periodic perturbation term which can be defined consistently with equation (2.5). For $\varepsilon = 0$, the system is left unperturbed, exhibiting a stable limit cycle dynamics on $\mathbf{x}(t) = ([S], [P])$ with the oscillation frequency $\omega_o(= 2\pi/T_o)$ (figure 1c).

For $\nu_0 = 0.005$ mM s^{-1}, $k_d = 0.05$ s^{-1} and $\varepsilon = 0$ (the yellow star in the phase diagram depicted in figure 1b), the substrate and product concentrations ($[S](t)$, $[P](t)$) undergo oscillation around the (unstable) fixed point $([S^*], [P^*]) = (0.22$ mM, 0.10 mM) (see the phase portrait in figure 1c), and this dynamics can be described using time-dependent phase angle and amplitude defined as $\theta(t) = \tan^{-1}(\delta[P](t)/\delta[S](t)) \in [0,2\pi)$ and $R(t) = \sqrt{(\delta[S](t))^2 + (\delta[P](t))^2}$, where $\delta[S] = [S] - [S^*]$ and $\delta[P] = [P] - [P^*]$. While the phase angle $\theta(t)$ is 2π-periodic, its variation is still non-uniform in time, i.e. $d\theta(t)/dt \neq$ const., as suggested by the varying size of velocity field in figure 1c. Thus, one can consider uniformizing the phase angle by defining a new phase angle $\phi(\theta)$,

$$\phi(\theta) = \omega_o \int_0^\theta \left(\frac{d\theta}{dt}\right)^{-1} d\theta, \quad (3.2)$$

which changes linearly in time for a limit cycle dynamics and satisfies $\phi(\theta + 2\pi) = \phi(\theta) + 2\pi$.
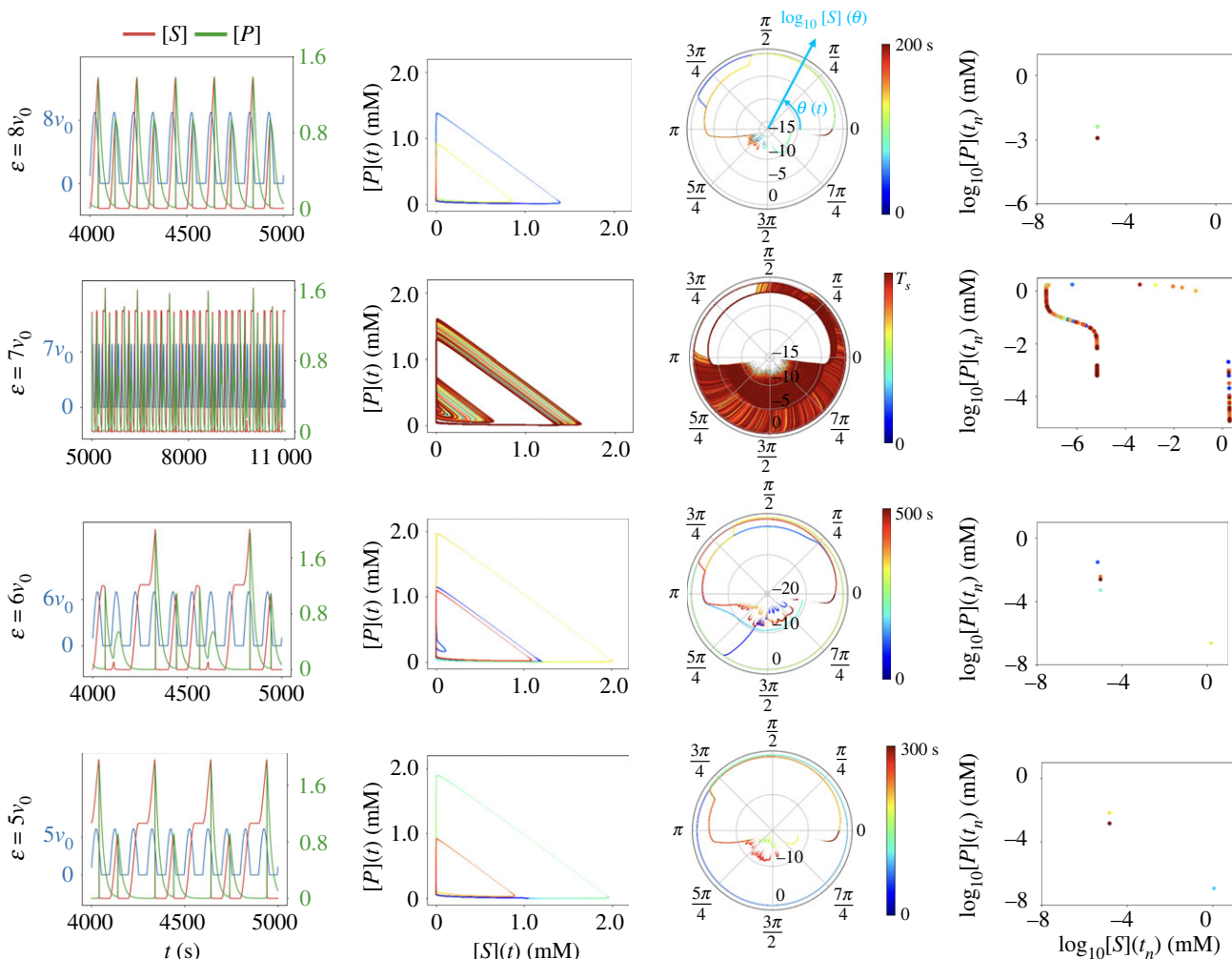
Figure 5. Trajectories generated under the conditions marked with four arrows in different colours in figure 4. The dynamical state that exhibits a period-3 cycle at $\varepsilon = 5\nu_0$ changes to a chaotic state at $\varepsilon = 7\nu_0$, and to a state with period-2 cycle at $\varepsilon = 8\nu_0$. The time-evolutions of $[S](t)$ (red) and $[P](t)$ (green) generated in response to $\nu(t)$ (blue) on the first column, and the plots of $[S](t)$ versus $[P](t)$ are shown on the second column. On the third column, the steady state trajectory of substrate concentration $[S](\theta)$ is depicted as a function of phase angle $\theta = \omega_{ext}t \mod 2\pi$ on the polar coordinate. The rightmost panels show the intersections of dynamic time trace across the Poincaré section.

Under a T_{ext}-periodic driving, $u(t + T_{ext}) = u(t)$ with small ε, one can formally recast equation (3.1) to an equation of motion in continuous time for the phase angle ϕ,

$$\frac{d\phi}{dt} = \frac{\partial\phi}{\partial[S]}\frac{d[S]}{dt} + \frac{\partial\phi}{\partial[P]}\frac{d[P]}{dt} \approx \omega_o + \varepsilon Q(\phi, t), \quad (3.3)$$

where $Q(\phi, t) = (\partial\phi/\partial[S])u(t)$. Integrated over a single period of driving T_{ext}, i.e. from t_n to t_{n+1}, where $t_n = 2\pi n/\omega_{ext}$, equation (3.3) can be written as a one-dimensional mapping from ϕ_n to ϕ_{n+1}, which corresponds to the *circle map* with an additional perturbation,

$$\phi_{n+1} = [\phi_n + 2\pi\Omega + \varepsilon R(\phi_n; \omega_{ext})] \mod 2\pi, \quad (3.4)$$

where $\phi_n = \phi(t_n)$, $\Omega = \omega_o/\omega_{ext} = T_{ext}/T_o$ and $R(\phi_n; \omega_{ext}) = \int_0^{T_{ext}} Q(\phi, t)\,dt$.

As $Q(\phi, t)$ is a 2π-periodic function of ϕ and a T_{ext}-periodic function of time t, the phase space of equation (3.3) is defined on a two-dimensional torus of $0 \leq \phi < 2\pi$ and $0 \leq t < T_{ext}$. For $\varepsilon = 0$, the dynamics of the uniformized phase angle forms a limit cycle with frequency ω_o, satisfying $\phi(t) = \omega_o t + \phi_0$. Importantly, there exists a one-to-one correspondence between the points on the unperturbed trajectory $\mathbf{x}_o(t) = ([S](t), [P](t))$ and the phase angle $\phi(t)$, such that $\mathbf{x}_o = \mathbf{x}_o(\phi)$ is an invertible function of ϕ. A forced dynamical system with $\varepsilon \neq 0$

still sustains its oscillatory trajectory $\mathbf{x}_\varepsilon(\phi)$ around the stable limit cycle $\mathbf{x}_o(\phi)$ as long as ε remains small. However, when ε is too large, $\mathbf{x}_\varepsilon = \mathbf{x}_\varepsilon(\phi)$ becomes non-invertible, so that the one-to-one mapping between $\mathbf{x}_\varepsilon$ and ϕ no longer holds, which leads to disruption of the torus [37]. In the case of the *sine circle map*, $R(\phi_n)$ in equation (3.4) is given as $R(\phi_n) = \sin\phi_n$, and hence the map becomes non-invertible if $\partial_\phi(\phi + 2\pi\Omega + \varepsilon\sin\phi) = 1 + \varepsilon\cos\phi \geq 0$ for some ϕ, which is realized when $\varepsilon \geq 1$. In our PFK model of glycolytic oscillation, it is found that the transition to non-invertibility, which permits the formation of chaotic orbits, is made for $\varepsilon \gtrsim 0.8\nu_0$ (figure 2a).

4. Discussions

The actual glycolytic oscillation is far more complicated than the simplified model explored here [8,11,12,38–40]. However, the overall dynamical responses of the system to the external periodic driving, are recapitulated as (i) the harmonic and sub/super-harmonic entrainments inside the Arnold tongues at weak-to-moderate driving, (ii) quasi-periodic states outside the Arnold tongues, and (iii) emergence of chaotic behaviours at strong driving. These are generic features expected for a dynamical system demonstrating a self-sustained oscillation [36] and

should neither be limited to the model of glycolytic oscillation nor depend on its microscopic details. Despite the diversity of network designs for biochemical oscillations, one can still use the general notions of dynamical system to gain reasonable understanding of their responses to periodic drivings.

Whereas this study discusses the emergence of chaotic behaviours from the conventional PFK model of glycolysis under non-autonomous external driving, chaotic dynamics can also arise autonomously for a more complex version of glycolysis. Decroly & Goldbeter [41] have studied another PFK model of glycolysis with three variables constituting two positive feedback loops, showing that the model, albeit autonomous, exhibits chaotic dynamics as well as both simple and birhythmic oscillations.

In order for a yeast cell to display glycolytic oscillations, the glucose injection rate along with the product degradation rate must be in a proper range. The phase diagram of PFK model built under the condition of constant glucose injection rate (figure 1b) suggests that an increased glucose injection rate surpassing $v_0 \approx 0.01$ mM s^{-1} ($k = 0.05$ s^{-1}) results in the cessation of glycolytic oscillation, which could imply a physiological state potentially associated with the hyperglycaemia. This study, however, suggests that the dynamical response of the system exhibiting a limit cycle to the external forcing is extremely rich. As long as the glucose is injected in a time-varying (periodic) manner over a proper range of frequency and amplitude, the system can be entrained into the Arnold tongues, still sustaining its oscillatory dynamics around its natural frequency. Although the role of glycolytic oscillations in physiology is not fully elucidated, glycolytic oscillations have been suggested as a pacemaker of other physiological oscillations [42–44]. Specifically, an oscillation of NADH, a product of glycolysis and the precursor of Kreb/electron transfer cycle, drives the oscillation of mitochondrial membrane potential [45]. Sustained oscillations in lactate released from islets of Langerhans provide a mechanism for pulsatile insulin secretion from β-cells [5]. Furthermore, acetaldehyde, a membrane-permeating metabolite from glycolysis, mediates cell-to-cell communication among a yeast cell population leading to in-phase oscillations [46–50].

The enhancement of the harmonic and sub/super-harmonic entrainments of glycolytic oscillation under the weak-to-medium periodic drivings (*broadening of the entrainment bands*) could be discussed from a more general perspective considering a phase angle dynamics under non-autonomous time-varying perturbation expressed in the form of equation (3.3). Specifically, recent studies on the synchronization among phase oscillators coupled with a coupling strength γ that can be mapped to ε in the present study, have also shown enlarged Arnold tongues with increasing γ [51], indicating the enhanced stability of oscillatory dynamics or *chronotaxicity*, in other words the ability of a self-sustained oscillator to resist non-autonomous perturbations. Along with these studies [51,52], our study substantiates that the glycolytic oscillation of a single cell can indeed be influenced by the oscillatory metabolic signals from the surrounding cells, and explains the ease of intercellular synchronization and enhanced stability of oscillations in cell populations [50,53,54]. However, an excessive amount of glucose injection can bring the system into disarray, rendering the dynamical state of the system chaotic, in other words, hyper-sensitive to initial conditions and unpredictable.

5. Methods

5.1. Evaluation of Lyapunov exponents

For general d-dimensional continuous-time dynamical equation satisfying a set of ODEs with the vector function $\mathbf{F}[\mathbf{x}(t)]$ over the d-dimensional vector $\mathbf{x}(t)$,

$$\frac{d\mathbf{x}(t)}{dt} = \mathbf{F}[\mathbf{x}(t)], \tag{5.1}$$

its variation of variables, $\mathbf{x} \to \mathbf{x} + \delta\mathbf{x}$, yields

$$\frac{d\delta\mathbf{x}(t)}{dt} = \mathbf{F}[\mathbf{x}(t) + \delta\mathbf{x}(t)] - \mathbf{F}[\mathbf{x}(t)] \approx \frac{\partial\mathbf{F}}{\partial\mathbf{x}} \cdot \delta\mathbf{x}(t), \tag{5.2}$$

for $\delta\mathbf{x}(t) \to 0$, and it formally satisfies

$$\delta\mathbf{x}(t) = \mathbf{M}(t, \mathbf{x}(0)) \cdot \delta\mathbf{x}(0), \tag{5.3}$$

where $\mathbf{M}(t, \mathbf{x}(0)) \equiv T_\leftarrow \exp\int_0^t \frac{\partial\mathbf{F}(\mathbf{x}(\tau))}{\partial\mathbf{x}(\tau)} d\tau$ denotes an $d \times d$ time-ordered exponential operator. The Lyapunov exponent of a dynamical system dictates the growth rate of the size of the variation. Thus, one can consider

$$\|\delta\mathbf{x}(t)\|^2 = \delta\mathbf{x}(0)^T \cdot \|\mathbf{M}(t, \mathbf{x}(0)\|^2 \cdot \delta\mathbf{x}(0), \tag{5.4}$$

with

$$\|\mathbf{M}(t, \mathbf{x}(0))\|^2 = \sum_i \mathbf{u}_i(t, \mathbf{x}(0))\chi_i(t, \mathbf{x}(0)) \cdot \mathbf{u}_i(t, \mathbf{x}(0))^T,$$

where $\mathbf{u}_i(t, \mathbf{x}(0))$ and $\chi_i(t, \mathbf{x}(0))$ are the ith eigenvector and eigenvalue of $\|\mathbf{M}(t, \mathbf{x}(0))\|^2$ matrix, respectively. The Lyapunov exponent associated with the change in the vector size $\|\delta\mathbf{x}(t)\| \sim e^{\lambda t}\|\delta\mathbf{x}(0)\|$ is calculated as follows [55]:

$$\lambda(\mathbf{x}(0)) = \lim_{t\to\infty} \frac{1}{2t} \ln(\chi_{\max}(t, \mathbf{x}(0))), \tag{5.5}$$

where $\chi_{\max}$ is the largest eigenvalue among the set of eigenvalues $\{\chi_i\}$ of $\|\mathbf{M}(t, \mathbf{x}(0))\|^2$ matrix.

For a practical numerical procedure to evaluate the Lyapunov exponent, we used the one proposed by Benettin *et al.* [56]. We consider d possible variations from d-dimensional vector $\mathbf{x}(0)$, say, $\delta\mathbf{x}_1(0), \delta\mathbf{x}_2(0), \ldots, \delta\mathbf{x}_d(0)$. Since these variations are not generally orthonormal to each other, the Gram–Schmidt procedure is used to obtain a corresponding set of orthonormal vectors $\delta\hat{\mathbf{X}}(0) = \{\delta\hat{\mathbf{x}}_1(0), \delta\hat{\mathbf{x}}_2(0), \ldots, \delta\hat{\mathbf{x}}_d(0)\}$, which span the same subspace by $\delta\mathbf{X}(0) = \{\delta\mathbf{x}_1(0), \delta\mathbf{x}_2(0), \ldots, \delta\mathbf{x}_d(0)\}$. Specifically, we build mutually orthogonal vectors

$$\mathbf{w}_k(0) = \delta\mathbf{x}_k(0) - \sum_{i=1}^{k-1} \langle\delta\mathbf{x}_k(0), \delta\hat{\mathbf{x}}_i(0)\rangle\delta\hat{\mathbf{x}}_i, \tag{5.6}$$

and normalize them using

$$\delta\hat{\mathbf{x}}_k(0) = \frac{\mathbf{w}_k(0)}{\|\mathbf{w}_k(0)\|}. \tag{5.7}$$

We integrate equation (5.1) to calculate the next position $\mathbf{x}(\tau)$ and employ equation (5.3) to calculate the next variations $\delta\mathbf{X}(\tau) \equiv [\delta\mathbf{x}_1(\tau), \delta\mathbf{x}_2(\tau), \ldots, \delta\mathbf{x}_d(\tau)]$. We then again apply the Gram–Schmidt procedure to obtain the corresponding set of orthonormal vectors $\delta\hat{\mathbf{x}}_1(\tau), \delta\hat{\mathbf{x}}_2(\tau), \ldots, \delta\hat{\mathbf{x}}_d(\tau)$. We repeat this procedure of integration and orthonormalization over the time duration of $N\tau$ for the full time trace.

At time $\tau \gg 1$, we expect

$$\|\delta\mathbf{x}_k(\tau)\| \sim e^{\lambda_k\tau}\|\delta\hat{\mathbf{x}}_k(0)\|. \tag{5.8}$$

Hence, the Lyapunov exponent corresponding to the growth rate of the variation in the kth direction is given by

$$\lambda_k = \lim_{\tau\gg 1} \frac{1}{\tau} \ln\frac{\|\delta\mathbf{x}_k(\tau)\|}{\|\delta\hat{\mathbf{x}}_k(0)\|}. \tag{5.9}$$

This can be cast into a matrix form

$$\delta \mathbf{X}(\tau) = \mathbf{M}(\tau, \mathbf{x}(0)) \cdot \delta \hat{\mathbf{X}}(0) = e^{\Lambda \tau} \cdot \delta \hat{\mathbf{X}}(0), \qquad (5.10)$$

where $\mathbf{M}(\tau, \mathbf{x}(0))$, $\delta \mathbf{X}(0)$, and $\Lambda = (\lambda_1, \ldots, \lambda_d) \cdot \mathbb{I}$ are $d \times d$ matrices, and it yields

$$\|\det \mathbf{M}(\tau, \mathbf{x}(0))\| = \exp\left[\left(\sum_{k=1}^{d} \lambda_k\right)\tau\right]. \qquad (5.11)$$

Therefore,

$$\sum_{k=1}^{d} \lambda_k = \lim_{\tau \gg 1} \frac{1}{\tau} \ln \|\det \mathbf{M}(\tau, \mathbf{x}(0))\|$$

$$= \lim_{\tau \gg 1} \frac{1}{\tau} \sum_{k=1}^{n} \ln \|\mathbf{w}_k(\tau)\|. \qquad (5.12)$$

Note that the transformation $\mathbf{M}(\tau, \mathbf{x}(0)) : \delta \hat{\mathbf{X}}(0) \to \delta \mathbf{X}(\tau)$ maps the d-dimensional *hypercube* of volume 1 formed by the d orthonormal vectors of variation $\delta \hat{\mathbf{X}}(0)$ into an d-dimensional *parallelopiped* made of $\delta \mathbf{X}(\tau)$. Thus, (i) $\sum_k \lambda_k$ measures the growth rate of the d-dimensional volume from the hypercube to the parallelopiped. (ii) The second line of the equation (5.12) follows from the fact that the determinant of the operator $\mathbf{M}(\tau, \mathbf{x}(0))$ is identical to the volume of the d-dimensional parallelopiped, Vol(P), and this volume can be calculated using the product of mutually orthogonalized vectors of $\delta \mathbf{x}_k(\tau)$, i.e. $\mathbf{w}_k(\tau)$ (equation (5.6)). Taken together,

$$\|\det \mathbf{M}(\tau, \mathbf{x}(0))\| = \text{Vol}(P) = \prod_{k=1}^{d} \|\mathbf{w}_k(\tau)\|. \qquad (5.13)$$

In our study, we aim to decide the growth (or convergence) rate of the trajectory averaged over the period of the external driving along the kth variation. Provided that the time traces are generated over N iterations of the period at steady state, we set $\tau = T_{\text{ext}}$ and consider calculating the Lyapunov exponent averaged over the N times of driving period.

$$\lambda_k = \frac{1}{NT_{\text{ext}}} \sum_{i=1}^{N} \ln \|\mathbf{w}_k(iT_{\text{ext}})\|. \qquad (5.14)$$

Finally, the Lyapunov exponent characterizing the dynamical behaviour of the system is obtained from the set of $\{\lambda_1, \lambda_2, \ldots, \lambda_d\}$ with $\lambda_1 > \lambda_2 > \cdots > \lambda_d$, and the largest Lyapunov exponent λ_1 decides the dynamical behaviour of the system at a long time limit.

Our system subject to an external periodic driving (equation (3.1)) can be recast in the form of a set of ODEs, $\dot{\mathbf{x}} = \mathbf{F}(\mathbf{x})$, with three variables ($d = 3$), $\mathbf{x} = ([S], [P], \psi)$,

$$\left. \begin{aligned} \partial_t[S] &= F_1([S], [P]) + \varepsilon \sin \psi \\ \partial_t[P] &= F_2([S], [P]) \\ \partial_t \psi &= \omega_{\text{ext}}. \end{aligned} \right\} \qquad (5.15)$$

Since ψ is T_{ext}-periodic, the corresponding Lyapunov exponent is always 0. Hence, $\lambda_1 = 0$ and $\lambda_2, \lambda_3 < 0$ for the case of periodic or quasi-periodic orbits, whereas $\lambda_1 > 0$ for chaotic orbits. In the phase diagrams of Lyapunov exponents in figures $2a$ and $4a$, we report the value of exponent λ, such that we use the positive largest Lyapunov exponent ($\lambda = \lambda_1 > 0$) to quantify the growth rate of variation for chaotic states and the negative largest Lyapunov exponent ($\lambda = \lambda_2 < 0$) to quantify the convergence rate for periodic or quasi-periodic states:

$$\lambda = \begin{cases} \lambda_1 & \text{for } \lambda_1 > 0 \\ \lambda_2 & \text{for } \lambda_1 = 0. \end{cases} \qquad (5.16)$$

Appendix A

A.1. Effect of stochastic forcing on self-sustained oscillations

In comparison with the periodic forcing (equation (2.5)), the stochastic forcing can be written as $v(t) = v_0 + \xi(t)$ with $\langle \xi(t) \rangle = 0$ and $\langle \xi(t)\xi(\tau) \rangle = \varepsilon \delta(t - \tau)$. Boiteux *et al.* varied the glucose injection rate (30–180 mM h$^{-1} \simeq 8.33 \times 10^{-3} - 0.05$ mM s^{-1}) 'stochastically' around its average value $v_0 (= 0.005$ mM s^{-1}), finding that the system retains its autonomous oscillation around its natural frequency [4].

In the light of equation (3.3), their finding can be formulated into the following dynamics of a limit cycle in a phase variable ϕ: $d\phi/dt = \omega_0 + \xi(t)$. Thus, it follows that $\mu_\phi \equiv \langle \delta\phi(t) \rangle = \omega_0 t$ and $\sigma_\phi^2 \equiv \langle (\delta\phi(t))^2 \rangle = \varepsilon t$, where $\langle \cdots \rangle$ denotes the average over trajectories. If one were to have a relative error (noise-to-signal ratio) less than 1, i.e. $\sigma_\phi/\mu_\phi (= \sqrt{\varepsilon t}/\omega_0 t) < 1$, one obtains a condition of $t > \varepsilon/\omega_0^2$. This means that even when subject to a stochastic forcing with a strength ε, the underlying phase variable dynamics of a self-sustained oscillation with the frequency ω_0 can still be read out as long as the observation time t is greater than ε/ω_0^2.

References

1. Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Walter P. 2008 *Molecular biology of the cell*, 5th edn. New York, NY: Garland Science.

2. Duysens L, Amesz J. 1957 Fluorescence spectrophotometry of reduced phosphopyridine nucleotide in intact cells in the near-ultraviolet and visible region. *Biochim. Biophys. Acta* **24**, 19–26. (doi:10.1016/0006-3002(57)90141-5)

3. Hess B, Boiteux A. 1971 Oscillatory phenomena in biochemistry. *Annu. Rev. Biochem.* **40**, 237–258. (doi:10.1146/annurev.bi.40.070171.001321)

4. Boiteux A, Goldbeter A, Hess B. 1975 Control of oscillating glycolysis of yeast by stochastic, periodic, and steady source of substrate: a model and

experimental study. *Proc. Natl Acad. Sci. USA* **72**, 3829–3833. (doi:10.1073/pnas.72.10.3829)

5. Chou H, Berman N, Ipp E. 1992 Oscillations of lactate released from islets of Langerhans: evidence for oscillatory glycolysis in beta-cells. *Am. J. Physiol. Endocrinol. Metab.* **262**, E800–E805. (doi:10.1152/ajpendo.1992.262.6.E800)

6. Tornheim K. 1997 Are metabolic oscillations responsible for normal oscillatory insulin secretion? *Diabetes* **46**, 1375–1380. (doi:10.2337/diab.46.9.1375)

7. Bertram R, Satin L, Zhang M, Smolen P, Sherman A. 2004 Calcium and glycolysis mediate multiple bursting modes in pancreatic islets. *Biophys. J.* **87**, 3074–3087. (doi:10.1529/biophysj.104.049262)

8. Goldbeter A, Lefever R. 1972 Dissipative structures for an allosteric model. *Biophys. J.* **12**, 1302–1315. (doi:10.1016/S0006-3495(72)86164-2)

9. Danø S, Sørensen PG, Hynne F. 1999 Sustained oscillations in living cells. *Nature* **402**, 320–322.

10. Reijenga KA, Westerhoff HV, Kholodenko BN, Snoep JL. 2002 Control analysis for autonomously oscillating biochemical networks. *Biophys. J.* **82**, 99–108. (doi:10.1016/S0006-3495(02)75377-0)

11. Termonia Y, Ross J. 1981 Oscillations and control features in glycolysis: numerical analysis of a comprehensive model. *Proc. Natl Acad. Sci. USA* **78**, 2952–2956. (doi:10.1073/pnas.78.5.2952)

12. Madsen MF, Danø S, Sørensen PG. 2005 On the mechanisms of glycolytic oscillations in yeast. *FEBS J.* **272**, 2648–2660. (doi:10.1111/j.1742-4658.2005.04639.x)

13. Goldbeter A. 1973 Patterns of spatiotemporal organization in an allosteric enzyme model. *Proc. Natl Acad. Sci. USA* **70**, 3255–3259. (doi:10.1073/pnas.70.11.3255)

14. Hess B. 1979 The glycolytic oscillator. *J. Exp. Biol.* **81**, 7–14. (doi:10.1242/jeb.81.1.7)

15. Hess B, Boiteux A. 1971 Oscillatory phenomena in biochemistry. *Ann. Rev. Biochem.* **40**, 1237.

16. Strogatz SH. 2014 *Nonlinear dynamics and chaos: with applications to physics, biology, chemistry, and engineering*. Boulder, CO: Westview Press.

17. Pikovsky A, Rosenblum M, Kurths J. 2002 *Synchronization: a universal concept in nonlinear science*. Cambridge, UK: Cambridge University Press.

18. Ott E. 2002 *Chaos in dynamical systems*. Cambridge, UK: Cambridge University Press.

19. Sel'kov E. 1968 Self-oscillations in glycolysis 1. A simple kinetic model. *Eur. J. Biochem.* **4**, 79–86.

20. Monod J, Changeux J-P, Jacob F. 1963 Allosteric proteins and cellular control systems. *J. Mol. Biol.* **6**, 306–329. (doi:10.1016/S0022-2836(63)80091-1)

21. Monod J, Wyman J, Changeux JP. 1965 On the nature of allosteric transitions: a plausible model. *J. Mol. Biol.* **12**, 88–118. (doi:10.1016/S0022-2836(65)80285-6)

22. Changeux JP. 2012 Allostery and the Monod-Wyman-Changeux model after 50 years. *Annu. Rev. Biophys.* **41**, 103–133. (doi:10.1146/annurev-biophys-050511-102222)

23. Cui Q, Karplus M. 2008 Allostery and cooperativity revisited. *Protein Sci.* **17**, 1295–1307. (doi:10.1110/ps.03259908)

24. Motlagh HN, Wrabl JO, Li J, Hilser VJ. 2014 The ensemble nature of allostery. *Nature* **508**, 331–339. (doi:10.1038/nature13001)

25. Thirumalai D, Hyeon C, Zhuravlev PI, Lorimer GH. 2019 Symmetry, rigidity, and allosteric signaling: from monomeric proteins to molecular machines. *Chem. Rev.* **119**, 6788–6821. (doi:10.1021/acs.chemrev.8b00760)

26. Kim P, Hyeon C. 2021 Thermodynamic optimality of glycolytic oscillations. *J. Phys. Chem. B* **125**, 5740–5749. (doi:1021/acs.jpcb.1c01325)

27. McCoy MA, Senior MM, Wyss DF. 2005 Screening of protein kinases by ATP-STD NMR spectroscopy. *J. Am. Chem. Soc.* **127**, 7978–7979. (doi:10.1021/ja0425942)

28. Moyer M, Gilbert S, Johnson K. 1998 Pathway of ATP hydrolysis by monomeric and dimeric kinesin. *Biochemistry* **37**, 800–813. (doi:10.1021/bi9711184)

29. Blangy D, Buc H, Monod J. 1968 Kinetics of the allosteric interactions of phosphofructokinase from *Escherichia coli*. *J. Mol. Biol.* **31**, 13–35. (doi:10.1016/0022-2836(68)90051-X)

30. Cardon JW, Boyer PD. 1978 The rate of release of ATP from its complex with myosin. *Eur. J. Biochem.* **92**, 443–448. (doi:10.1111/j.1432-1033.1978.tb12765.x)

31. Gilbert SP, Johnson KA. 1994 Pre-steady-state kinetics of the microtubule-kinesin ATPase. *Biochemistry* **33**, 1951–1960. (doi:10.1021/bi00173a044)

32. Song Y, Hyeon C. 2021 Thermodynamic uncertainty relation to assess biological processes. *J. Chem. Phys.* **154**, 130901. (doi:10.1063/5.0043671)

33. Arnold VI. 1965 Small denominators, I: mappings of the circumference into itself. *AMS Trans. Series 2* **46**, 213. (doi:10.1007/BF00275153)

34. Jensen MH, Bak P, Bohr T. 1983 Complete devil's staircase, fractal dimension, and universality of mode-locking structure in the circle map. *Phys. Rev. Lett.* **50**, 1637–1639. (doi:10.1103/PhysRevLett.50.1637)

35. Bak P. 1986 The devil's staircase. *Phys. Today* **39**, 38–45. (doi:10.1063/1.881047)

36. Heltberg ML, Krishna S, Kadanoff LP, Jensen MH. 2021 A tale of two rhythms: locked clocks and chaos in biology. *Cell Syst.* **12**, 291–303. (doi:10.1016/j.cels.2021.03.003)

37. Zhusubaliyev ZT, Mosekilde E. 2009 Novel routes to chaos through torus breakdown in non-invertible maps. *Physica D* **238**, 589–602. (doi:10.1016/j.physd.2008.12.012)

38. Smolen P. 1995 A model for glycolytic oscillations based on skeletal muscle phosphofructokinase kinetics. *J. Theor. Biol.* **174**, 137–148. (doi:10.1006/jtbi.1995.0087)

39. Bier M, Teusink B, Kholodenko BN, Westerhoff HV. 1996 Control analysis of glycolytic oscillations. *Biophys. Chem.* **62**, 15–24. (doi:10.1016/S0301-4622(96)02195-3)

40. Chandra FA, Buzi G, Doyle JC. 2011 Glycolytic oscillations and limits on robust efficiency. *Science* **333**, 187–192. (doi:10.1126/science.1200705)

41. Decroly O, Goldbeter A. 1982 Birhythmicity, chaos, and other patterns of temporal self-organization in a multiply regulated biochemical system. *Proc. Natl Acad. Sci. USA* **79**, 6917–6921. (doi:10.1073/pnas.79.22.6917)

42. Rapp P. 1987 Why are so many biological systems periodic? *Prog. Neurobiol.* **29**, 261–273. (doi:10.1016/0301-0082(87)90023-2)

43. Xiong L, Garfinkel A. 2023 Are physiological oscillations *physiological*? *J. Physiol.* (doi:10.1113/JP285015)

44. Langlhofer G, Kogel A, Schaefer M. 2021 Glucose-induced $[Ca^{2+}]_i$ oscillations in β cells are composed of trains of spikes within a subplasmalemmal microdomain. *Cell Calcium* **99**, 102469. (doi:10.1016/j.ceca.2021.102469)

45. Olsen LF, Andersen AZ, Lunding A, Brasen JC, Poulsen AK. 2009 Regulation of glycolytic oscillations by mitochondrial and plasma membrane H^+-ATPases. *Biophys. J.* **96**, 3850–3861. (doi:10.1016/j.bpj.2009.02.026)

46. Richard P, Bakker BM, Teusink B, Van Dam K, Westerhoff HV. 1996 Acetaldehyde mediates the synchronization of sustained glycolytic oscillations in populations of yeast cells. *Eur. J. Biochem.* **235**, 238–241. (doi:10.1111/j.1432-1033.1996.00238.x)

47. Bier M, Bakker BM, Westerhoff HV. 2000 How yeast cells synchronize their glycolytic oscillations: a perturbation analytic treatment. *Biophys. J.* **78**, 1087–1093. (doi:10.1016/S0006-3495(00)76667-7)

48. Palková Z, Váchová L. 2006 Life within a community: benefit to yeast long-term survival. *FEMS Microbiol. Rev.* **30**, 806–824. (doi:10.1111/j.1574-6976.2006.00034.x)

49. Weber A, Zuschratter W, Hauser MJ. 2020 Partial synchronisation of glycolytic oscillations in yeast cell populations. *Sci. Rep.* **10**, 19714. (doi:10.1038/s41598-020-76242-8)

50. De Monte S, d'Ovidio F, Danø S, Sørensen PG. 2007 Dynamical quorum sensing: population density encoded in cellular dynamics. *Proc. Natl Acad. Sci. USA* **104**, 18 377–18 381 (doi:10.1073/pnas.0706089104)

51. Lucas M, Newman J, Stefanovska A. 2018 Stabilization of dynamics of oscillatory systems by nonautonomous perturbation. *Phys. Rev. E* **97**, 042209. (doi:10.1103/PhysRevE.97.042209)

52. Lancaster G, Suprunenko YF, Jenkins K, Stefanovska A. 2016 Modelling chronotaxicity of cellular energy metabolism to facilitate the identification of altered metabolic states. *Sci. Rep.* **6**, 29584. (doi:10.1038/srep29584)

53. Aon MA, Cortassa S, Lemar KM, Hayes AJ, Lloyd D. 2007 Single and cell population respiratory oscillations in yeast: a 2-photon scanning laser microscopy study. *FEBS Lett.* **581**, 8.14 (doi:10.1016/j.febslet.2006.11.068)

54. Hauser MJ. 2022 Synchronisation of glycolytic activity in yeast cells. *Curr. Genet.* **68**, 69–81. (doi:10.1007/s00294-021-01214-y)

55. Dorfman JR. 1999 *An introduction to chaos in nonequilibrium statistical mechanics*, vol. 14. Cambridge, UK: Cambridge University Press.

56. Benettin G, Galgani L, Giorgilli A, Strelcyn J-M. 1980 Lyapunov characteristic exponents for smooth dynamical systems and for hamiltonian systems; a method for computing all of them. Part 2: numerical application. *Meccanica* **15**, 21–30. (doi:10.1007/BF02128237)