# Cell volume cell stiffness and cell motion

Dave Weitz Harvard

NIH, NSF, Harvard MRSEC

Ming Guo Allen Ehrlicher Karen Kasza Fred MacKintosh Angelo Mao

Harvard	Dave Mooney	Harvard
McGill	Enhua Zhou	HSPH
Sloan-Kett.	Jeff Fredberg	HSPH
Amsterdam	Jennifer Lippincott-Schwartz	NIH
Harvard	Tommy Angelini	Florida

- 1. Bulk osmotic modulus of cells
- 2. Substrate stiffness and cell volume
- 3. Dynamic arrest in motion of cells

http://weitzlab.seas.harvard.edu/

Active Systems GIST, 6/27/14

#### Cell stiffness measurements

**(B)** 





Frequency (Hz)

trap and bead displacements (nm)

# Control of cell volume

- What controls cell volume
- What are consequences of cell volume

# Bulk modulus (osmotic)

- Susceptibility to change in volume
- Not due to compression of water
- Due to change in volume of cell

## Compression of cell volume with PEG



Cell volume responds immediately to change in osmotic pressure

#### Cell volume controlled by osmotic pressure (PEG)



One single cell being compressed repeatedly.



# Control of cell volume

- Water eflux through membrane
- Depends on osmotic pressure

# П-V relationship



# П-V relationship



# Control of cell volume

- Water eflux through membrane
- Depends on osmotic pressure
- Stresses on membrane ~ shear modulus
- Too weak to support pressure difference
- No pressure drop across membrane

# $\rightarrow$ Pressure inside = pressure outside



 $\Pi_{\rm osm}$ 

(Pa)

ions, while  $V_{\min}$  is occupied by cellular materials, mainly proteins.

#### Substrate stiffness dependence of equation of state



#### Number of intracellular osmolytes



The concentration of osmolyte estimated is on the order of  $10^2$  mM; the number agrees with intracellular ions measured experimentally.

# Bulk (osmotic) modulus of cells



- Bulk modulus *B* >> shear modulus *G*
- Cytoskeleton plays negligible role

Cell volume is strongly regulated → What are the consequences

### Cells have many different stiffnesses



#### Cell stiffness depends on stiffness of environment



Cell Stiffness:



#### Stem-cell fate depends on stiffness of environment



Cell Stiffness:



#### Cells adjust stiffness to their environments

Cell stiffness changes when growing on adhesive patterns with various sizes.



Tee et al. Biophysical J (2011)

Cell stiffness changes with osmolarity.



Zhou et al. PNAS (2009)

Why does cell stiffness change? What control parameters are involved?

# Cell morphology

## Cell area changes with substrate stiffness



#### Projected area of cell increases with substrate stiffness.

### Cell area increases with substrate stiffness





Projected cell area increases with increasing substrate stiffness.

### Cell height decreases with substrate stiffness



While the projected cell area increases with increasing substrate stiffness, the cell height decreases.

### Cell height decreases with substrate stiffness



# Discrete slice through cell

### Cell height decreases with substrate stiffness







While the projected cell area increases with increasing substrate stiffness, the cell height decreases.

#### Cell volume decreases with substrate stiffness



### A7 Cell size distribution



~200 cells per distribution

# Control cell spreading by adherence area



Contact printing of adhesion proteins

# Control cell spreading by adherence area



Cells spread to cover only printed area

# Control cell spreading by adherence area



10 µm

# Cell volume decreases with area



## Dependence of cell volume on cell area



Control adhesive area

# Dynamics of cell volume changes

Spreading of a single cell



# Dynamics of cell volume changes



# Dynamics of stiffness adjustment



### Effect of molecular motors


### Depends on water content of cell



## Cell stiffness depends on substrate stiffness





### Cell stiffness depends on substrate stiffness









Behaves like reconstituted cross-linked actin networks:  $C^{2.2}$ 

#### Cell stiffness scaling is same as that of reconstituted actin network



# Cortical and cytoplasmic stiffness scales with cell volume



## Nucleus volume changes too



## Nucleus volume changes with cell volume



# What about stem-cell differentiation?

D1 mouse mesenchymal stem cells on stiff and soft PAA gel substrates (collagen coated)

Compress cell volume osmotically with adding PEG 300.



Bar: standard error \* p < 0.05

 $_{\rm or}$  Soft substrate  $\rightarrow$  volume decreased by osmotic pressure

### Cell volume affects stem cell differentiation



Bar: standard error \* p < 0.05

Soft substrate  $\rightarrow$  volume decreased by osmotic pressure

# Full gene expression analysis

Gene-chip analysisAnalysis of mRNA

• Determine origin of dependence

- Cell stiffness correlates with cell volume
- Cell volume controllable by:
  - substrate stiffness
  - adhesion area
  - osmotic pressure
- Nucleus volume tracks cell volume
  - gene expression
  - stem-cell differentiation

# Now for something fun

# Cell motion on substrates

# Single cell versus multi-cell motion





#### **Tissue cells:**

- contractile "inchworm"
- spread, stick, release

# Single cell versus multi-cell motion



30 µm

#### **Tissue cells:**

- contractile "inchworm"
- spread, stick, release



30 *µ*m

Do canonical forms drive collective motion?

# Q: how does a fibroblast talk to his friends a millimeter away?

## How do $10^5 - 10^6$ cells coordinate?



Trepat, et. al., Nat. Phys. 2009

#### Do cells communicate across millimeters?



**tissue-cell collective migration**: development, wound healing, tumor invasion

MDCK epithelial cells on polyacrylamide surfaces.

# Time evolution of cell behavior



#### MDCK epithelial cells on polyacrylamide surfaces.

Speed up time-lapse: cells move, substrate fluctuates.
200 mins, ~800 µm field of view. 16 datasets taken in succession.

# Cell density increases with time

200 mins, ~800  $\mu$ m field of view. 16 datasets taken in succession.



# Traction force microscopy



### MDCK epithelial cells on polyacrylamide surfaces.



#### Cells

#### **Substrate**

#### Deformation patterns span hundreds of microns



#### 800 microns

Quantify length-scale: spatial correlation function

$$C_{dd}(R) = \left\langle \frac{\sum_{j} \delta \mathbf{d}(\mathbf{r}_{j}) \cdot \delta \mathbf{d}(\mathbf{r}_{j} + \mathbf{R})}{\sum_{j} \delta \mathbf{d}(\mathbf{r}_{j}) \cdot \delta \mathbf{d}(\mathbf{r}_{j})} \right\rangle_{t,\varphi}$$

### Correlation length grows in time



# Collective migration: large scale patterns



$$C_{\mathbf{vv}}(R) = \left\langle \frac{\sum_{j} \delta \mathbf{v}(\mathbf{r}_{j}) \cdot \delta \mathbf{v}(\mathbf{r}_{j} + \mathbf{R})}{\sum_{j} \delta \mathbf{v}(\mathbf{r}_{j}) \cdot \delta \mathbf{v}(\mathbf{r}_{j})} \right\rangle_{t,\varphi}$$

### Measure correlation length



## Correlation correlation length grows in time



### How are the two patterns related?



### Swirl size grows with deformation pattern



### Migration follows deformation in time



$$C_{\mathbf{dv}}(\tau) = \left\langle \sum_{j} \delta \mathbf{d}(t_{j}) \cdot \delta \mathbf{v}(t_{j} + \tau) \right\rangle_{\mathbf{R}}$$

- •Time correlation: positive slope.
  - •Slope decreases with time.

# Substrate deformations: essential







Cells live on a substrate that is roughly as stiff as they are 200 + 1 Cells call long distance: collective substrate deformations guide collective cell migration



Dynamic structure factor

 $S(q,\omega) = \left| \mathscr{F}[\rho(r,t)] \right|^2$ 

Sound speed, c
Damping, Γ
Diffusivity, D<sub>0</sub>
Compressibility, χ
DOS

# $S(q,\omega)$ of cell motion pick a q, analyze $\omega$ lineshape


# $\begin{aligned} & \text{Rayleigh-Brillouin triplet} \\ & \frac{S(q,\omega)}{S(q)} = \frac{I_R(q)\frac{1}{2}\Gamma_0(q)}{\omega^2 + \left(\frac{1}{2}\Gamma_0(q)\right)^2} + \frac{I_B(q)\Omega(q)\Gamma^2(q)}{\left(\omega^2 - \Omega^2(q)\right)^2 + \omega^2\Gamma^2(q)} \end{aligned}$



•Commonly used function in IXS, INS

•**Rayleigh** peak: diffusive fluctuations.

•**Brillouin** peak pressure fluctuations.

Vogel - Fulcher Tammann Hesse (VFTH) equation



Assume  $D_{i} = (cell size)^{2}/z_{bus}$  VFTH fit gives  $\pi_{p} = 2728 \text{ mm}^{-2}$ colloidal glass transition

#### Divergence of time scale for colloidal particles



Bartch, Antonietti, Schupp, Sillescu, J. Chem. Phys. 97, 3950 (1992)



# $\frac{S(q,\omega)}{S(q)} = \frac{I_R(q)\frac{1}{2}\Gamma_0(q)}{\omega^2 + \left(\frac{1}{2}\Gamma_0(q)\right)^2} + \frac{I_B(q)\Omega(q)\Gamma^2(q)}{\left(\omega^2 - \Omega^2(q)\right)^2 + \omega^2\Gamma^2(q)}$



•Commonly used function in IXS, INS

•**Rayleigh** peak: diffusive fluctuations.

•Brillouin peak pressure fluctuations.

#### Brillouin peak



#### Density of States (DOS)



#### Brillouin peak: Density of States (DOS)



#### Brillouin peak: Density of States (DOS)



DOS peak: soft modes in glasses

#### Velocity field: length scale for flow



#### Structural Relaxations in a Supercooled Fluid



#### Relaxing particles are highly correlated spatially

#### Relaxation events are spatially correlated



volume fraction

Cluster size grows as glass transition is approached

## Dynamic heterogeneities grow

choose all v's within fastest 15% calculate size of connected regions:  $\xi_h$ 



Migration slows down, cluster size grows. Analog to dynamic heterogenities in colloidal glasses.

# Dynamic heterogeneities grow





### Compressibility



Wavelength at peak = **0.40+-0.04 cell lengths**. Exactly the same as **high-***\omega* **DOS peak**! Phase transition from **migration** state to **division** state?

### Tissue analogy to glass forming fluid

- Growing dynamic heterogeneities
- Growing relaxation times: fragility
- Cell body deformation: Boson peak in DOS

Implications:

- collective motion is activated.
- fluidize tissue to initiate fast motion.
- wound healing, metastasis, embryonic development.

## Conclusions

- •Inert particles undergo 'diffusive' motion
- •Fluctuations driven by motors in cell
- •Cell volume depends on cell spreading
- •Cell stiffness correlates with cell volume
- •Stem-cell differentiation depends on volume

## Thank you for your attention