











Rule 2: Don't believe the protocol of anyone who is not a motility expert



Growing in glucose gives few flagella Everyday pipette tips/filtration will shear off flagella Corollary: if in doubt (and even if not), look!









Section I:Theoretical background

I. Revision (low Re theory)

2. Introduction to swimming micro-organisms

3. Real vs. model E. coli

4. Detailed theory of model *E. coli*









$$\rho a = -\frac{dP}{dx} + \eta \frac{d^2 u}{dz^2} + f$$

characteristic speed and length: U, L
Dimensionless variables:
 $\bar{x}, \bar{z} = \frac{x}{L}, \frac{z}{L}$
 $\bar{u} = \frac{u}{U}$
 $\bar{a} = \frac{a}{U^2/L}$
 $\bar{p} = \frac{P}{\eta U/L}$
 $\bar{f} = \frac{f}{\eta U/L^2}$
Re $\eta \frac{d\bar{P}}{d\bar{x}} + \frac{d^2\bar{u}}{d\bar{z}^2} + \bar{f} = 0$
 $-\nabla P + \eta \nabla^2 \mathbf{v} + \mathbf{f} = 0$





































$$\mathbf{F}(t) = \mathbf{F}_{\text{head}}(t) + \int_{0}^{L} ds \mathbf{f}(s, t) = 0 . \tag{6}$$






















Linda Turner











$$\begin{pmatrix} \mathbb{F}_{b} \\ \mathbb{N}_{b} \end{pmatrix} = -\begin{pmatrix} A_{0} & 0 \\ 0 & D_{0} \end{pmatrix} \begin{pmatrix} \mathbf{v} \\ \mathbf{\Omega} \end{pmatrix} \text{ and} \\ \begin{pmatrix} \mathbb{F}_{f} \\ \mathbb{N}_{f} \end{pmatrix} = -\begin{pmatrix} A & B \\ B & D \end{pmatrix} \begin{pmatrix} \mathbf{v} \\ \boldsymbol{\omega} \end{pmatrix}, \text{ with} \\ A_{0}, D_{0}, A, B, D > \mathbf{0} \\ \mathbf{v} = (v, 0, 0), \, \boldsymbol{\omega} = (-\omega, 0, 0), \, \boldsymbol{\Omega} = (\Omega, 0, 0) \\ \text{Force free } : -A_{0}v - Av - B(-\omega) = 0 \text{ or } (A_{0} + A)v = B\omega \\ \text{Torque free } : -D_{0}\Omega - Bv - D(-\omega) = 0 \text{ or } D_{0}\Omega = -Bv + D\omega \\ \text{(Neglects hydrodynamic interaction between body and flagella!)} \end{cases}$$











Force free
$$: -A_0v - Av - B(-\omega) = 0$$
 or $(A_0 + A)v = B\omega$
Torque free $: -D_0\Omega - Bv - D(-\omega) = 0$ or $D_0\Omega = -Bv + D\omega$
 $M_m = \Omega + \omega = \Omega + \beta\Omega = (1 + \beta)\Omega$
 $M_m = \beta(A_0, D_0, A, B, D)$
 $M_m = D_0\Omega$
Completely specifies model *E. coli*, provided A₀, D₀, A, B, D are known











$$\mathbf{v}_{SS} = \frac{F}{8\pi\eta} \left(\frac{x^2 + r^2}{r^3}, \frac{xy}{r^3}, \frac{xz}{r^3} \right) \qquad \mathbf{v}_{SD} = \frac{G}{4\pi} \left(\frac{1}{r^3} - \frac{3x^2}{r^5}, -\frac{3xy}{r^5}, -\frac{3xz}{r^5} \right)$$
$$G = \frac{Fa^2}{6\eta}$$
$$\mathbf{v}_{SS}(r = a) + \mathbf{v}_{SD}(r = a) = \frac{F}{6\pi\eta a} (1, 0, 0)$$
Satisfies boundary condition for sphere translating with $\mathbf{v} = (u, 0, 0)$ Sphere exerts force $\mathbf{F} = 6\pi\eta a \mathbf{v}$ on fluid
Fluid exerts drag $\mathbb{F} = -6\pi\eta a \mathbf{v}$ on sphere
Indeed $\mathbf{F} \propto \mathbf{v}$ (linearity)





$$A = \xi_{\perp} \ell \frac{1 - \beta}{\sqrt{\beta}} \left(1 + \gamma_k \frac{\beta}{1 - \beta} \right), \qquad \gamma_k = \xi_{\parallel} / \xi_{\perp}$$

$$B = \xi_{\perp} \ell \left(\frac{\lambda}{2\pi} \right) \frac{1 - \beta}{\sqrt{\beta}} (1 - \gamma_k), \qquad \xi_{\perp} (1 - \gamma_k) = \xi_{\perp} - \xi_{\parallel}$$

$$D = \xi_{\perp} \ell \left(\frac{\lambda}{2\pi} \right)^2 \left(1 + \gamma_k \frac{\beta}{1 - \beta} \right) \frac{1 - \beta}{\sqrt{\beta}}$$

$$\beta = \cos^2 \psi \text{ with } \psi = \tan^{-1} (2\pi R / \lambda) \text{ is the helix pitch angle}$$
















Chattopadhyay et al. *PNAS* 103 (2013) 13712–13717

We use measurements of swimming bacteria in an optical trap to determine fundamental properties of bacterial propulsion. In particular, we directly measure the force required to hold the bacterium in the optical trap and determine the propulsion matrix, which relates the translational and angular velocity of the flagellum to the torques and forces propelling the bacterium.













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On Torque and Tumbling in Swimming Escherichia coli[⊽]†

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Bacteria swim by rotating long thin helical filaments, each driven at its base by a reversible rotary motor. When the motors of peritrichous cells turn counterclockwise (CCW), their filaments form bundles that drive the cells forward. We imaged fluorescently labeled cells of *Escherichia coli* with a high-speed charge-coupled-device camera (500 frames/s) and measured swimming speeds, rotation rates of cell bodies, and rotation rates of flagellar bundles. Using cells stuck to glass, we studied individual filaments, stopping their rotation by exposing the cells to high-intensity light. From these measurements we calculated approximate values for bundle torque and thrust and body torque and drag, and we estimated the filament stiffness. For both immobilized and swimming cells, the motor torque, as estimated using resistive force theory, was significantly lower than the motor torque reported previously. Also, <u>a bundle of several flagella produced little more torque than a single flagellum produced. Motors driving individual filaments frequently changed directions of rotation. Usually, but not always, this led to a change in the handedness of the filament, which went through a sequence of polymorphic transformations, from normal to semicoiled to curly 1 and then, when the motor again spun CCW, back to normal. Motor reversals were necessary, although not always sufficient, to cause changes in filament chirality. Polymorphic transformations among helices having the same handedness</u>

Single 'effective flagellum' model remains ill tested ...

... may be necessarily non-self-consistent!

Other ways of testing Stokes propulsion in Newtonian fluids ...

