

Unexpected Swelling of Stiff DNA in a Polydisperse Crowded Environment

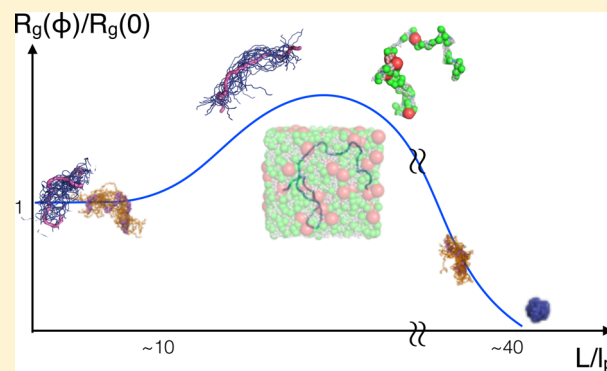
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S Supporting Information

ABSTRACT: We investigate the conformations of DNA-like stiff chains, characterized by contour length (L) and persistence length (l_p), in a variety of crowded environments containing monodisperse soft spherical (SS) and spherocylindrical (SC) particles, a mixture of SS and SC, and a milieu mimicking the composition of proteins in the *Escherichia coli* cytoplasm. The stiff chain, whose size modestly increases in SS crowders up to $\phi \approx 0.1$, is considerably more compact at low volume fractions ($\phi \leq 0.2$) in monodisperse SC particles than in a medium containing SS particles. A 1:1 mixture of SS and SC crowders induces greater chain compaction than the pure SS or SC crowders at the same ϕ , with the effect being highly nonadditive. We also discover a counterintuitive result that the polydisperse crowding environment, mimicking the composition of a cell lysate, swells the DNA-like polymer, which is in stark contrast to the size reduction of flexible polymers in the same milieu. Trapping of the stiff chain in a fluctuating tube-like environment created by large-sized crowders explains the dramatic increase in size and persistence length of the stiff chain. In the polydisperse medium, mimicking the cellular environment, the size of the DNA (or related RNA) is determined by L/l_p . At low L/l_p , the size of the polymer is unaffected, whereas there is a dramatic swelling at an intermediate value of L/l_p . We use these results to provide insights into recent experiments on crowding effects on RNA and also make testable predictions.



INTRODUCTION

The recognition that the crowded cellular environment can profoundly influence all biological processes, such as gene expression,^{1–3} protein^{4–6} and RNA folding,^{7–9} and protein–protein interactions,¹⁰ has received increasing attention recently, although its importance was recognized decades ago.¹¹ A simple calculation using the typical concentration of macromolecules in the *Escherichia coli* shows that the average spacing between proteins is ~ 4 nm, which is comparable to the size (radius of gyration) $R_g \approx 0.3N^{1/3}$ nm of a folded protein¹² with $N \approx 300$ amino acid residues. Therefore, the cellular interior, replete with macromolecules of different sizes and shapes (a polydisperse soup), is crowded, affecting the stability and shapes of the molecules of life. For example, compaction of DNA, relevant in a variety of biological processes ranging from organization of the nucleoid in bacteria to DNA packaging in phage heads, is greatly facilitated in the presence of neutral osmotic agents. The effect of neutral polymer (polyethylene oxide) (PEO) in compacting DNA (Ψ condensation) was demonstrated in a pioneering study by Lerman,¹¹ who showed that DNA undergoes a dramatic reduction in size if the concentration of PEO exceeds a critical value. Because the interactions between DNA and PEO in these experiments was

established to be repulsive,¹³ Ψ condensation is determined solely by the volume excluded to DNA by the crowding particles. Subsequently, Post and Zimm¹⁴ produced insightful theoretical explanations based on further experiments on crowding-induced compaction of DNA. In the intervening years, a number of theoretical and experimental studies have explored various aspects of DNA compaction.^{15–17}

Despite these advances, a molecular description of how macromolecular crowding, especially a milieu containing polydisperse crowders, affects the spatial organization of DNA is poorly understood. Simulations of DNA in the presence of explicit crowding particles are computationally intensive, but their importance in describing the shapes of flexible polymers has been demonstrated in a number of recent studies.^{18–22} Because of technical complications, theories do not take into account the effects of polydispersity.^{15,16,23–30} In addition, the extent of structural changes and how they depend on mixtures of crowding particles of different shapes for realistic sizes of crowders found in the cytoplasm are unknown. Most of the studies have focused on intrinsically flexible

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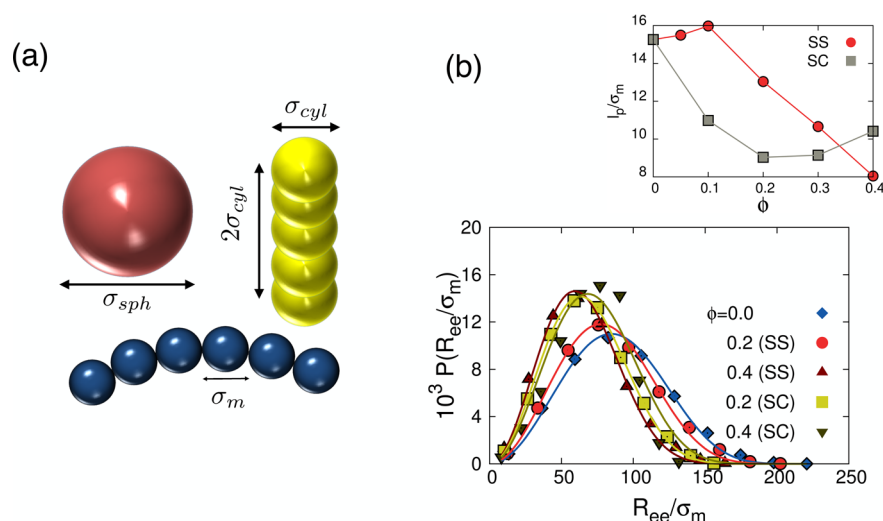


Figure 1. (a) Coarse-grained models of spherical (SS) crowder (red), spherocylindrical (SC) crowder (yellow), and WLC chain (blue) used to model DNA. The relevant dimensions are labeled σ_{sph} , σ_{cyl} , and σ_m . (b) End-to-end distance distribution of WLC at two different volume fractions ($\phi = 0.2, 0.4$) of SS and SC crowders. The top panel shows the persistence length (l_p) of the polymer as a function of the volume fraction of SS and SC crowders.

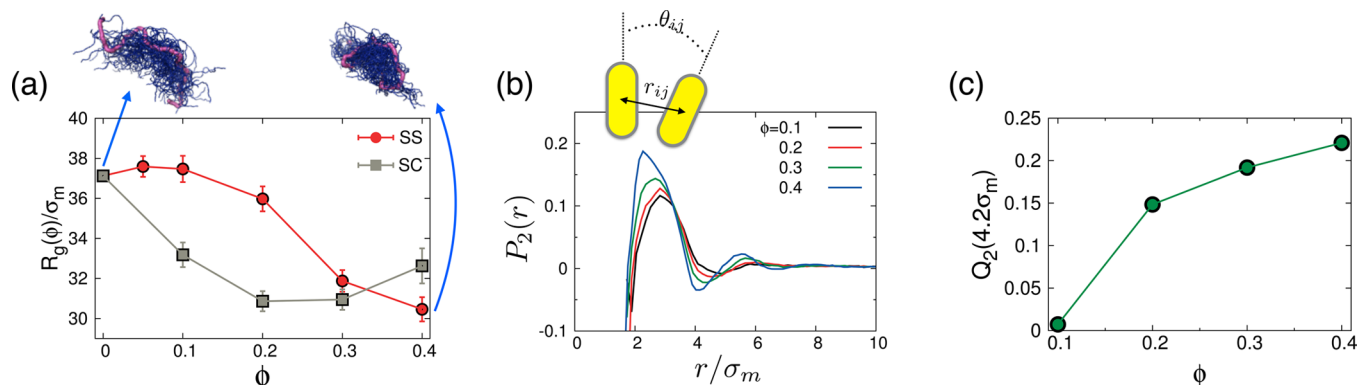


Figure 2. (a) Change of R_g of the polymer from $R_g(0) = 37.6$ nm with increasing ϕ . (b) Liquid crystal order parameter for SC crowders, averaged over the ensemble of crowder particles, as a function of distance. (c) Extent of local nematic ordering is quantified with $Q_2(r)$ at $r = 4.2\sigma_m$.

polymers with very little work on relatively short stiff DNA-like polymers and related RNA. Stiff short chains can display behavior quantitatively different than that of the associated long flexible polymers, as argued in the case of cyclization of DNA.^{31–33} These issues take on added importance because of potential relevance to genome confinement,^{4,34,35} where the structural organization is functionally related to gene expression.

Inspired by these observations, we first performed simulations of worm-like chain³⁶ (WLC, a reliable polymer model for describing many of the properties of DNA at high salt concentration) in monodisperse solutions containing soft spherical (SS) and spherocylindrical (SC) crowding particles. Here, we focus on DNA-like chains in the limit where the contour length ($L = N_m l_0$, where N_m is the number of monomers and l_0 is the bond length) is not significantly longer than the persistence length, l_p . The systematic study leads to a number of unexpected predictions, which can be tested using synthetic polymers in the presence of nanoparticles and DNA using crowding agents. (i) The SC crowders induce greater compaction than SS particles. The compaction is accompanied by substantial reduction of l_p (by nearly a factor of 2), similar in magnitude to that observed by ion-induced compaction of

DNA and RNA.³⁷ (ii) One of the most striking results of our study is that stiff chains are more compact in a mixture of SS and SC, at physiologically relevant volume fractions, than in monodisperse crowders at the same volume fraction. The substantial compaction of DNA in the mixture is due to ordering (dense packing) of the SS crowders around DNA due to depletion attraction induced by the SCs. (iii) We also carried out simulations of DNA in polydisperse crowders mimicking the composition of macromolecules in *E. coli*. Surprisingly, we obtained a counterintuitive and theoretically unanticipated result that polydisperse spherical crowding particles cause swelling of DNA at a volume fraction of ≈ 0.3 appropriate for *E. coli* cytoplasm. Surrounded predominantly by large-sized crowders, the swollen conformation of DNA (nearly 3-fold increase in volume of the chain) is entropically driven.

RESULTS

Crowding-Induced Softening of DNA. In order to set the scale for bending energy of the polymer, we first calculated the end-to-end distance distribution, $P(R_{ee})$, in the absence of crowders to obtain the bare persistence length, l_p , for $\phi = 0$. By fitting the simulated $P(r)$ to an analytic expression for WLC (eq S3),³¹ we obtained $l_p = (15.4 \pm 0.08)\sigma_m \approx 49$ nm using $\sigma_m \approx$

3.18 nm (Figure 1b), which coincides with l_p for DNA in high monovalent salt concentration. Because $l_p/\sigma_m > 1$, the shape of the chain in the presence of crowding particles should be determined by an interplay of bending rigidity and attractive depletion interaction due to crowders. $P(R_{ee})$ of the WLC with varying ϕ for both monodisperse SS and SC crowders shows a gradual shift to the smaller R_{ee} with increasing ϕ from 0 to 0.4 (Figure 1b). At high ϕ , as ϕ increases, l_p obtained from the fit of $P(r)$ to eq S3 gradually decreases for both SS and SC crowders, implying that the crowding particles induce compaction of the WLC polymer (Figure 1b, top panel). Interestingly, the WLC polymer exhibits a nonmonotonic dependence of size with increasing ϕ . The SS crowders induce a minor increase of l_p (stiffening or expansion) of the chain (Figure 1b top panel and Figure 2a) for $0 < \phi \leq 0.1$, followed by a decrease of l_p (softening or compaction) at larger $\phi = 0.2$ –0.4. In contrast, the SC crowders reduces l_p much more efficiently than the SS crowders at $\phi \leq 0.3$ in that an expansion similar to the one in the SS crowders at $\phi \approx 0$ –0.1 is not observed. Instead, there is a modest restiffening of the DNA due to SC crowders when ϕ is in the range from $\phi = 0.2$ to $\phi = 0.4$. We further substantiate this result below by calculating the change in the polymer size (R_g) and nematic order parameter of the crowders. The slight increase in l_p notwithstanding, the overall trend is that there is substantial softening (l_p decreases by nearly a factor of 2) as ϕ increases from 0 to 0.4.

Dependence of R_g on ϕ for Monodisperse Crowders.

Snapshots of polymer conformations at different values of ϕ show modest compaction as ϕ increases (Figure 2a). Similar to l_p , the dependence of R_g on ϕ for SS and SC crowders displays substantial difference. At $\phi = 0.2$, $R_g(0.2)$ is smaller than $R_g(0)$ by only 4% in SS crowders, whereas $R_g(0.2)$ decreases by 17% in SC crowders. Given that the volume of the chain is $\sim R_g^3(\phi)$, the extent of compaction induced by SC is substantial compared to that of SS crowders. The quantitative difference between the effects of SS and SC crowders on the chain compaction is explained using the depletion interaction (or Asakura–Oosawa (AO) interaction³⁸) that produces an effective attraction between monomers. The strength of the AO interaction for SS, roughly given by $\approx \phi k_B T / \sigma_{sph}^2$, has to exceed the energy ($\sim 1/2(L/l_p)k_B T$) required to bend the polymer on scale l_p for compaction to occur. For small ϕ , it is unlikely that the AO attraction can compensate for the bending penalty. Thus, we expect little change in $R_g(\phi)$ at small ϕ for SS crowders. On the other hand, the strength of the AO interaction on the WLC for SC crowders is $\approx \phi P(\sigma_m/\sigma_{cyl}^2) k_B T$, where $P(=2\sigma_{cyl})$ is the cylinder length. In both cases, the origin of the AO depletion interaction, which has to exceed the bending energy to compact the stiff chain, leading to an effective short-range (on length scale of $\sim \sigma_m$) attraction between monomers and polymer compaction, is purely entropic. Figure 2a shows that $R_g(\phi)$ decreases monotonically until $\phi \approx 0.3$ for SC, indicating that, for the parameter used, the strength of the attractive AO interaction due to SC crowders can exceed the penalty for bending the chain on the scale of $\sim l_p$.

The strong effect of compaction of the WLC chain induced by SC relative to SS crowders can also be quantified by comparing the volume excluded to the polymer by the crowders. On the basis of scaled particle theory, we can estimate the entropy cost of inserting a hard sphere of dimension σ_{HS} in a box containing hard fluid particles. The

entropy difference for inserting the hard sphere of diameter σ_{HS} is related to $C(\sigma)$ ³⁹

$$C(\sigma_{HS}) = \frac{V_{cyl}}{V_{sph}} = \frac{3[(\sigma_{HS} + \sigma_{cyl})^2 P + (\sigma_{HS} + \sigma_{cyl})^3]}{2(\sigma_{HS} + \sigma_{sph})^3} \quad (1)$$

where V_{cyl} (V_{sph}) is the volume excluded by rod-like (spherical) crowders. For the parameters listed in Table S1, we find that $V_{cyl} > V_{sph}$ provided $\sigma_{HS} \approx R_g > \sigma_{cyl}$ or σ_{sph} . The entropic cost of inserting a spherical particle of size R_g into a fluid of cylindrical crowders exceeds that for inserting it into a system consisting of spherical crowders. By achieving greater compaction of the WLC in SC crowders, the entropy difference is minimized, thus explaining the results in Figure 2a.

The pattern of compaction in the two crowding environments is qualitatively different. For SS crowders, compaction occurs only when ϕ exceeds ≈ 0.1 ($dR_g/d\phi \approx 0$ as $\phi \rightarrow 0$). Although the trend is not so clear as in $l_p(\phi)$ (Figure 1b, top panel), $R_g(\phi)$ (Figure 2a) shows a signature of minor swelling in the range of $\phi = 0$ –0.1, which was also observed in experiments on the effects of crowding on a ribozyme (see below).⁸ In contrast, for anisotropic crowding agents (SC crowders), $R_g(\phi)$ decreases monotonically for $\phi \leq 0.3$ ($dR_g/d\phi < 0$ for all ϕ) and increases from $\phi = 0.3$ to $\phi = 0.4$. High volume fraction of SC results in the reswelling of WLC (Figure 2a), showing that the shape of the crowding particles can have a profound effect on the size of a stiff chain.

Local Nematic Order and Increase in R_g . Interestingly, as ϕ exceeds 0.3, $R_g(\phi)$ of the chain increases in SC crowders (Figure 2a). The increase in $R_g(\phi)$ at higher ϕ is due to plausible development of local nematic ordering for SC crowders at $\phi > 0.3$. The system consisting of pure SC crowders will undergo an isotropic to nematic phase transition if ϕ exceeds a critical value $\phi_{I \rightarrow N}$. We calculated the liquid crystal order parameter, $\langle P_2(\cos \theta) \rangle$, where θ is the angle between the long axes of any pairs of SC crowders, $P_2(x) = 1/2(3x^2 - 1)$ is the second Legendre polynomial, and $\langle \dots \rangle$ denotes the ensemble average. $\langle P_2(\cos \theta) \rangle$ is almost zero (< 0.05) for all ϕ , which means that even the highest ϕ ($= 0.4$) is less than $\phi_{I \rightarrow N}$. However, locally, the crowders adopt a nematic-like state. To ascertain if this is the case, we calculated $P_2(r) = \langle \sum_{i,j} P_2(\cos \theta_{ij}) \delta(|\vec{r}_i - \vec{r}_j| - r) \rangle$ as a function of distance, r . Figure 2b shows that the angular correlation of SCs becomes stronger as ϕ increases at short distances.

To quantify the extent of “local” nematic ordering, we calculated $Q_2(r) = N_p^{-1} \int_{r_{min}}^r P_2(r') dr'$, where N_p is the number of pairs separated by r and r_{min} is the minimal distance between SC crowders. At the distance $r = 2.3\sigma_m$, where the pair correlation has a first peak for $\phi = 0.4$ (Figure 2b), $P_2(2.3\sigma_m)$ increases significantly from 0.0 to 0.23 as ϕ increases from 0.1 to 0.4 (Figure 2c). At $r = 4.2\sigma_m$, $Q_2(r) \approx 0.23$ suggests that the local nematic ordering of cylindrical crowders is reinforced in the vicinity of the WLC. This result implies that a relatively stiff chain induces ordering of SC crowders along the polymer axis and strengthens the anisotropic interaction of rod-like particles. At high ϕ , the chain may be thought of as being in a local nematic field, which elongates the polymer along the local direction of the nematic field, thus explaining the increase in $R_g(\phi)$ when ϕ exceeds 0.3.

Nonadditive Effect in a Mixture of Spheres and Spherocylinders. The dependence of $R_g(\phi)$ on ϕ of the

WLC in the 1:1 mixture of SS and SC crowders is shown in Figure 3. The mixture has a profound effect on the size of

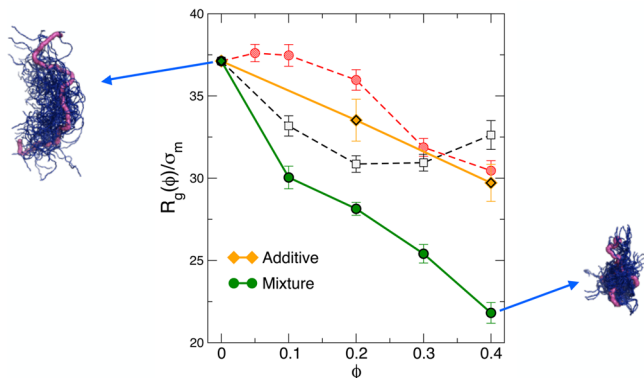


Figure 3. Compaction of a stiff chain in a mixture of SS and SC crowders. R_g of WLC as a function of ϕ of 1:1 SS and SC mixture (green). The orange line is the calculated R_g at each ϕ value by assuming that the effects of SS and SC crowders on WLC compaction are additive. The results of monodisperse SS and SC crowders in Figure 2a are shown with the dashed lines to underscore the substantial enhancement of the chain compaction by the mixture.

DNA. The value of $R_g(0.4)$ is reduced by over 40% from $R_g(0)$, whereas the maximum compaction in monodisperse SC at $\phi = 0.4$ is only 10%. In order to highlight the nonadditive effects of the two crowders with different shape, we also show, at $\phi = 0.2$ and $\phi = 0.4$, the expected result for $R_g^A(\phi) = R_g(0) + \delta R_g^A(\phi)$, where $\delta R_g^A(\phi) = \delta R_g^{SS}(\phi/2) + \delta R_g^{SC}(\phi/2)$, obtained using the assumption that the effect of crowders on the extent of R_g reduction is additive. Remarkably, $R_g(\phi)$ is significantly lower than $R_g^A(\phi)$ (Figure 3) (or $|\delta R_g(\phi)| > |\delta R_g^A(\phi)|$, $|\delta R_g^{SC}(\phi)|$, $|\delta R_g^{SS}(\phi)|$), indicating that the mixture of SS and SC restricts the volume available to the WLC to a much greater extent than the individual components do.

The surprising finding of significant compaction in the mixture can be qualitatively explained using the notion of depletion potentials for mixture of SS and SC. Consider the interaction between two spherical particles in the presence of rods (SC particles). If a SC particle is spatially trapped, then the rod loses translational and rotational entropy because of orientational restrictions. The large unfavorable entropy loss results in the depletion of the SC from the space where the SC particle is trapped. The result is that there would be an excess osmotic pressure due to the AO attraction that pushes the SS and the monomers together. Two consequences of the entropy-

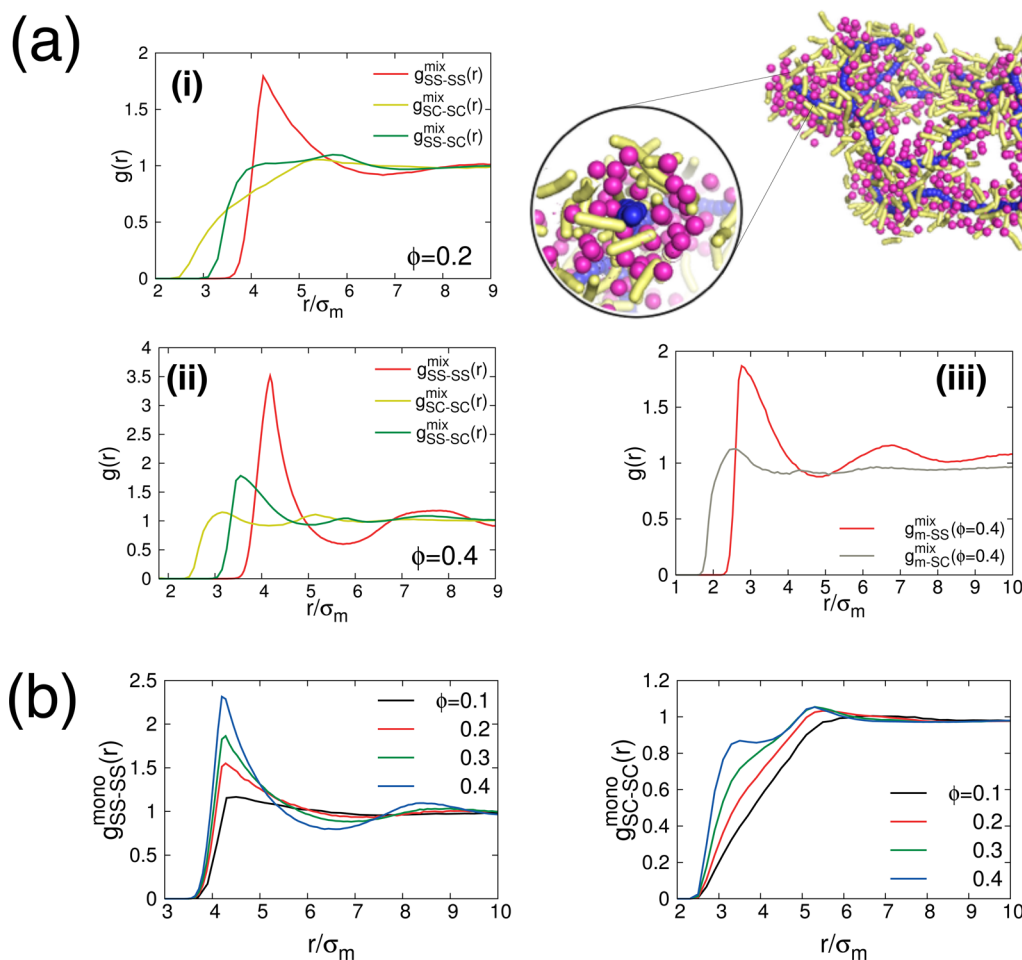


Figure 4. (a) RDFs are calculated for all possible combinations of crowder–crowder pairs at (i) $\phi = 0.2$ and (ii) 0.4 and (iii) monomer crowder (at $\phi = 0.4$). Distribution of SS and SC crowders around the WLC chain is depicted using a snapshot from the simulations (the particle sizes are not to scale). Although $\sigma_{\text{sph}} = 4\sigma_m$ and $\sigma_{\text{cyl}} = 4^{2/3}\sigma_m$ were used in actual simulations, we deliberately reduced the sizes of SS and SC crowders for clear illustration of the crowders around the chain. (b) RDFs of SS–SS and SC–SC pairs at $\phi = 0.1$ – 0.4 in monodisperse conditions are presented to highlight the enhanced local ordering of SS and SC crowders in the mixture.

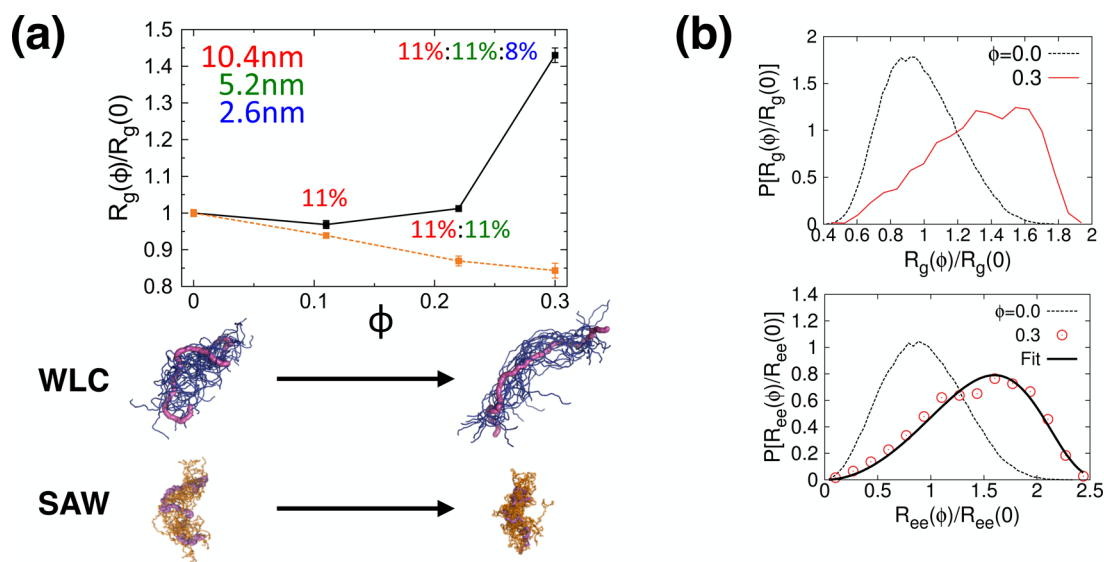


Figure 5. Effects of polydisperse soft sphere mixture on the size of WLC ($L/l_p = 20$ at $\phi = 0$). (a) R_g is calculated for WLC (black solid line) and SAW (orange dashed line) polymers at (i) $\phi = 0.11$ of crowders with $r_1 = 10.4$ nm, (ii) $\phi = 0.22$ with $r_1 = 10.4$ nm, $r_2 = 5.2$ nm (11%), and (iii) $\phi = 0.3$ with $r_1 = 10.4$ nm (11%), $r_2 = 5.2$ nm (11%), $r_3 = 2.6$ nm (8%). The largest crowder represents the ribosomes; the 5.2 nm crowders correspond to polymerases and other large protein complexes, and the smallest-sized particles are the average size of other crowders in the milieu. Structural ensembles of polymers at $\phi = 0$ and 0.3 are shown at the bottom, demonstrating the contrasting effect of polydisperse crowders on the conformations of WLC and SAW. (b) $P(R_g)$ (top) and $P(R_{ee})$ (bottom) of WLC at $\phi = 0$ and 0.3. The fit of $P(R_{ee})$ at $\phi = 0.3$ to eq S3 in the text gives $l_p^{\phi=0.3} = 41\sigma_m \approx 130$ nm, which is ~ 2.6 -fold greater than $l_p^{\phi=0} (\approx 15\sigma_m \approx 49$ nm). $P(R_g)$ of SAW (flexible self-avoiding chain) is shown in Figure S3.

driven depletion interactions include the following. (i) Due to the attractive interactions, the SS particles are more closely packed than in the absence of the SC (compare $g_{SS-SS}^{\text{mix}}(r)$ at ϕ from Figure 4a and $g_{SS-SS}^{\text{mono}}(r)$ at $\phi/2$ from Figure 4b (monodisperse crowders)). Indeed, in the limit of $\sigma_{\text{cyl}} < P < \sigma_{\text{sph}}$, it has been experimentally shown that the addition of a small fraction (by volume) of the SC can even lead to crystallization of low density suspension of hard spheres.^{40,41} (ii) We also expect that the excess volume available to the WLC should be greatly reduced compared to the monodisperse crowders. In such a confined space, the WLC should be considerably more compact than in the presence of monodisperse crowders at the same ϕ .

The expected enhancement in the packing of the SS due to the SC is evident in the pair distribution function $g(r)$. The results for $\phi = 0.2$ and $\phi = 0.4$ (Figure 4a(i,ii)) show that both the radial distribution functions (RDFs), $g_{SC-SC}^{\text{mix}}(r)$ between SCs and $g_{SC-SS}^{\text{mix}}(r)$ between SC and SS, do not exhibit significant structure. In sharp contrast, $g_{SS-SS}^{\text{mix}}(r)$ has the structure corresponding to a high density liquid (especially at $\phi = 0.4$ in Figure 4a(ii)), which is remarkable given that at $\phi/2 (= \phi_{SS} = 0.2)$, corresponding to the same volume fraction occupied by SS, $g_{SS-SS}^{\text{mono}}(r)$ (Figure 4b) is relatively featureless.

The much stronger depletion force due to the SC crowders results in a considerable reduction in the volume accessible to the chain, which explains the dramatic reduction of $R_g(\phi)$ compared to the pure component case. The WLC at $\phi = 0.4$ is surrounded predominantly by SS crowders, which is shown by $g_{m-X}^{\text{mix}}(r)$, the RDF between the monomer and the crowders ($X = \text{SS}$ or SC) (Figure 4a(iii)). The number of SS near the monomer calculated using $N_{SS} = 4\pi \left(\frac{N_0}{V} \right) \int_0^{r_{\text{min}}} r^2 g_{m-SS}(r) dr$, where r_{min} is the first minimum in the $g_{m-SS}(r)$ at $\phi = 0.4$ and is 11.4. A similar calculation for SC gives $N_{SC} = 4.7$.

Polydisperse Crowders Mimicking the *E. coli* Cytoplasm Composition. To a first approximation, the *E. coli* cytoplasm may be represented by a mixture of spheres because a majority of the crowders present in large numbers (ribosomes, polymerases, and other large complexes and smaller particles) are compact.⁴² In order to assess the shape of a DNA-like chain in such a mixture, we investigated the effect of polydisperse spherical particles on the conformational fluctuations of the stiff polymer.

Strikingly, the behavior of the stiff chain in a polydisperse mixture of SS particles differs drastically from those in the mixture of SS and SC of the same volume (Figure 3). A few features in the nonmonotonic dependence of $R_g(\phi)$ as a function of ϕ (Figure 5a) are worth pointing out. (i) There is a very modest reduction in $R_g(\phi)$ at $\phi \approx 0.1$, which has only large crowding particles. Such a decrease is comparable to that found in Figure 2a. (ii) Unexpectedly, $R_g(\phi)$ starts increasing in a mixture containing large- and medium-sized crowders. Most strikingly, in the mixture roughly mimicking the *E. coli* cytoplasm,⁴² there is a large increase ($\sim 40\%$) in $R_g(\phi)$ compared to $\phi = 0$. The competition between bending free energy and depletion potential leading to a dramatic swelling of the stiff chain is counterintuitive. The ensemble of the chain conformations (Figure 5a) exhibiting the expansion of the chain captures these effects visually. (iii) The *E. coli* mixture dramatically stiffens the polymer. The persistence length of the chain for the swollen chain in the *E. coli* mixture is about 2.6 times larger than that for the one at $\phi = 0$. The stiffening effect of the mixture of spherical crowders on WLC captured the snapshots in Figure 5. (iv) It is noteworthy that in the polydisperse mixture with *E. coli* composition reduces the size of a flexible chain that lacks bending penalty (Figure 5a and Figure S3), underscoring the importance of chain stiffness.

The reswelling at high ϕ (Figure 5a,b) can be understood qualitatively using the following arguments. At a specified total

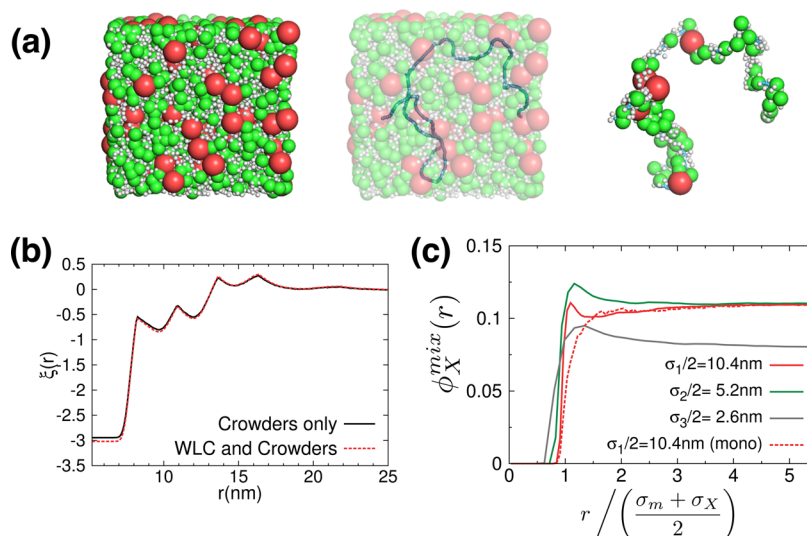


Figure 6. (a) Snapshot from simulation, demonstrating (left) the polydisperse crowding environment, (middle) the WLC inside the crowd, and (right) the crowding particles decorating the monomers. (b) Local size ordering correlation function, $\xi(r)$, indicates a nonuniform, heterogeneous size ordering of polydisperse crowd. (c) $\phi_X^{\text{mix}}(r)$ ($X = 1, 2, 3$), the fraction of volume around a monomer occupied by three different types of crowd in polydisperse crowding environment (case (iii) in (a)) as a function of monomer–crowd distance (solid lines). The dashed line shows the corresponding quantity for monodisperse crowding environment.

volume fraction, there are a lot more small crowding particles than large ones. Therefore, the entropy of the crowding particles is maximized if the WLC is surrounded by the larger-sized particles with the smaller ones being further away from the chain. In this picture, the WLC is localized in a region in which the larger-sized particles are with higher probability in proximity to the monomers (Figure S4). Because the interactions between the crowding particles and the stiff chain are repulsive, the DNA chain would prefer to be localized in a largely crowd-free environment. If we assume that such a region is roughly spherical, created predominantly by the largest crowd, then its size has to be on the order of R_g^3 to accommodate the WLC chain. In such a cavity, there is an entropic cost to confine the stiff chain. The probability of finding such a region decreases exponentially as R_g gets large. In addition, in a spherical region, the DNA would form spool-like structures requiring overcoming bending energy. The combination of these effects makes it likely that an optimal spherical regime can be found to minimize the free energy of WLC. If the region is cylindrical and large enough such that tight hairpins (costing substantial bending penalty) are avoided, then the chain free energy may be minimized by confining it in a roughly cylindrical cavity. Such a possibility is supported by simulations which show that, on average, the shape of the depletion zone is aspherical, resembling a fluctuating tube (see images in Figure 6a). As a result, we can visualize the polymer to be essentially confined to an anisotropic (but fluctuating) tube in which transverse fluctuations of the chain are restricted but one in which tight hairpin turns cannot form because of bending penalty. In such a cavity, the chain stiffens, thus expanding in size in order to minimize both the bending penalty and entropy cost of confinement. A more quantitative and accurate theory is difficult to construct because of the many-body correlation among the polydisperse crowding particles.

The plausibility of the physical picture given above is further substantiated by examining how the crowding particles with different sizes are arranged in space and how they surround the extended WLC. Distributions of the polydisperse crowding

particles are not uniform but exhibit local size ordering. This is evident in the local size correlation function (r -dependent size variance), $\xi(r) = \langle d_i d_j \rangle_r - \langle d \rangle^2$ (Figure 6b), where $\langle d \rangle$ is the mean diameter and $\langle d_i d_j \rangle_r$ in the first term denotes an average of the product of two diameters d_i and d_j taken over crowd (j) located at a distance r from a crowd i .⁴³ The local size correlation $\xi(r)$ (we set $\langle d_i d_j \rangle_r = 0$ when no pair exists), which is by definition zero for both monodisperse crowd and homogeneously distributed polydisperse crowd, reveals the presence of size ordering up to $r \leq 15\text{--}20\text{ nm}$. This implies that there is a nonuniform ordering in the mixture of spherical particles, which is responsible for reswelling (Figure 5a).

The fraction of volume occupied by three different crowd ($\phi_X^{\text{mix}}(r) = (4\pi/3) \times (\sigma_X/2)^3 \times \phi_{X\text{sgm-X}}^{\text{mix}}(r)$) with $X = 1, 2, 3$ as a function of distance from the WLC monomers in Figure 6c further captures the nonuniform distribution of crowd. The crowd with intermediate size ($X = 2$) occupy the largest volume near the WLC. In addition, the comparison of $\phi_2^{\text{mix}}(r)$ with $\phi_2^{\text{mono}}(r)$ (the volume fraction of monodisperse 11% crowd around WLC monomer; the dashed line in Figure 6c) shows that the intermediate ($X = 2$) and large-sized ($X = 1$) crowd are pushed closer to the monomers by the small crowd, which confines a segment of the polymer to a tube-like region (Figure 6a). The depletion forces in a polydisperse solution give rise to a spatial inhomogeneity of crowd, resulting in the chain being confined to a cylindrical region created by the large-sized ($X = 1, 2$) crowd. The expansion of the chain in such a confined space⁴⁴ provides a plausible physical explanation for the large increase in the size of the DNA.

DISCUSSION

Swelling and Collapse of DNA. The counterintuitive finding that a stiff polymer, with $\gamma = L/l_p$ not being large (see below), can swell in a polydisperse mixture of spheres is (to our knowledge) unprecedented. There are simulation and theoretical studies predicting the collapse of flexible polymers and proteins in mixed solvents due to volume exclusion effects

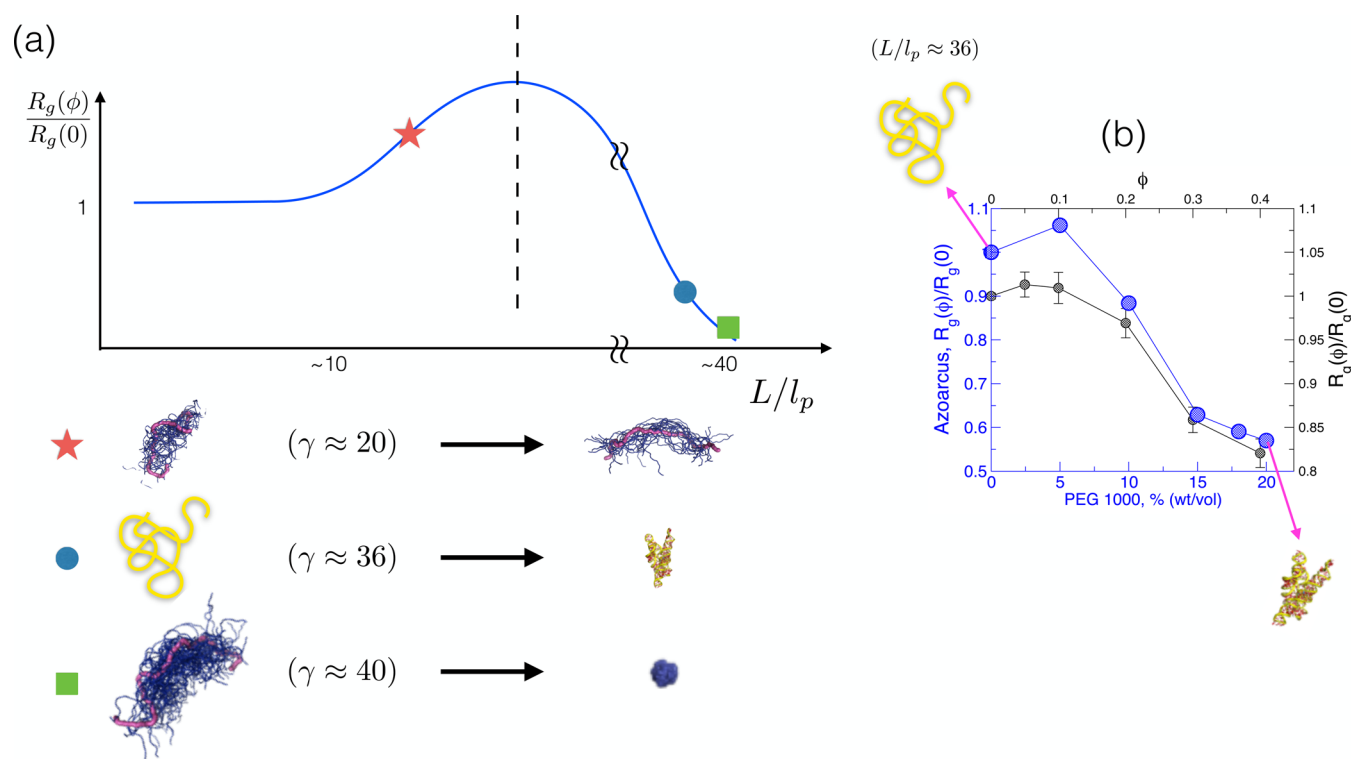


Figure 7. (a) Schematic of the expected changes in the size of biopolymers in the milieu of *E. coli*-like polydisperse crowding environment with $\phi = 0.3$ as L/l_p is varied. Depending on the parameter value $\gamma = L/l_p$, which characterizes the chain length and stiffness, the polymer undergoes swelling ($\gamma \geq 10$) or coil-to-globule transition ($\gamma \gg 10$). (b) PEG-induced compaction of *Azoarcus* ribozyme in 0.56 mM Mg^{2+} ion solution (blue circles). For comparison, we show in simulation (black circles) results for $R_g(\phi)$ changes in monodisperse SS crowding from Figure 2a.

alone.^{45,46} However, the present study shows precisely the opposite behavior for stiff chains, whereas a flexible chain tends to become compact (not a globule in the *E. coli* like milieu). We propose that this unusual effect is related to an interplay of chain bending and the complex depletion effect in a polydisperse crowding system. In order to substantiate our proposal, we carried out simulations for chains with γ varying from 2 to 10 in the model *E. coli*-like system. The simulations show hardly any change in R_g (see Figure S5). For all the values of γ (≤ 10), the stiff polymer can be accommodated in a large enough region in which the crowding particles do not suppress the conformational fluctuations. Only when γ exceeds a minimum value, but is not too large, does chain swelling occur by formation of tight turns.

We provide arguments that when γ exceeds a certain value, the DNA-like polymer must undergo a transition from the swollen state to a globule. In other words, there must be a sharp coil–globule transition induced by the crowders. When $\gamma \gg 1$, chain stiffness is not that relevant and the polymer behaves like a flexible polymer. Using our results in ref 19, we predict that γ has to exceed 40 to observe a genuine coil–globule transition. The simulation results and the physical arguments allow us to predict a rich dependence of R_g in the *E. coli* environment (Figure 7a).

Insights into Crowding Effects on RNA. Recent experiments have examined the effects of polyethylene glycol (PEG) on the folding of a ribozyme.⁸ It has been argued that the impact of PEG can be understood based on the excluded volume effect. The SAXS experiments on the *Azoarcus* ribozyme with 195 nucleotides shows that R_g , near the midpoint of the Mg^{2+} ion needed for the folding transition, initially increases before becoming compact (Figure 7b).

Folding of this RNA is also accompanied by decrease in the persistence length, which can be modulated by crowders. For *Azoarcus* ribozyme, $\gamma \approx 36$, where we have used $L \approx 195 \times 0.55$ nm and $l_p \approx 3$ nm.⁴⁷ If the theoretical prediction for the monodisperse SS in Figure 2a is correct, then we expect a modest increase in R_g as PEG (assumed to be spherical) concentrations increase. The experimental data are in qualitative agreement with this expectation. It would be most interesting to examine the effects of polydisperse crowding agents on the complex problem of RNA folding to further test some of our predictions.

CONCLUSIONS

In summary, using *explicit* simulations of crowding particles, we predict multiple and unexpected scenarios for the effects of a polydisperse crowding environment on the size and shape of a semiflexible polymer, which has served as a model for DNA and even RNA. Depending on the size, shape, and composition of the mixtures of crowding particles, we find evidence for both compaction and, surprisingly, dramatic increase in size, as well. The results are of great relevance to the recent explosion of interest in the behavior of RNA,⁸ DNA,^{26,48} and proteins^{49,50} in macromolecular crowding conditions both *in vitro* and *in vivo*. The prediction that shape of chains, such as DNA, RNA, and F-actin,⁵¹ can be dramatically altered in a polydisperse milieu can be tested in experiments.

METHODS

Model. To study the effects of crowding particles on a stiff chain, we used a coarse-grained model of WLC polymer ($N_m = 300$) and SS and SC crowding particles (Figure 1a). The length scales, σ_{sp}/l_p , σ_{cy}/l_p , and σ_m (≈ 3.18 nm for DNA) denote the size (diameter) of the SS, SC

crowding particles, and the monomer of WLC polymer, respectively. We set the aspect ratio of the SC to be 2 and $\sigma_{\text{sph}} = 2^{2/3}\sigma_{\text{cyl}} = 4\sigma_{\text{m}}$, so that the volumes of the individual SS and SC crowding particles are identical. In the WLC model, chain connectivity, with a fixed bond length, is maintained using a large spring constant connecting two consecutive beads. The bending rigidity of the chain was implemented by quadratic bond angle potential. We chose Weeks–Chandler–Andersen (WCA) potential for interactions between monomers, and r^{-12} soft sphere potential is employed for excluded volume interactions for crowder–crowder and crowder–polymer beads.

Soft Spheres. The energy function for the system consisting of the WLC and soft spherical crowders is

$$E = E_{\text{S}} + E_{\text{B}} + E_{\text{WCA}} + E_{\text{R}} \quad (2)$$

$$E_{\text{S}} = \sum_{i=1}^{N_{\text{m}}-1} K \frac{(|\vec{r}_{i+1} - \vec{r}_i| - l_0)^2}{l_0^2} \quad (3)$$

$$E_{\text{B}} = \sum_{i=1}^{N_{\text{m}}-1} G(\theta_i - \theta_0)^2 \quad (4)$$

$$E_{\text{WCA}} = \sum_{i,j < N_{\text{m}}} \Theta\left(\frac{\sigma_{ij}}{r_{ij}} - 1\right) \varepsilon \left[\left(\frac{\sigma_{ij}}{r_{ij}}\right)^{12} - \left(\frac{\sigma_{ij}}{r_{ij}}\right)^6 \right] \quad (5)$$

$$E_{\text{R}} = \sum_{i < j} \varepsilon \left(\frac{\sigma_{ij}}{r_{ij}}\right)^{12}, \quad \sigma_{ij} = \frac{\sigma_i + \sigma_j}{2} \quad (6)$$

where $\sigma_{ij} = (\sigma_i + \sigma_j)/2$, $\Theta(x)$ is a Heaviside step function, N_{m} is the number of monomers, and K ($>1000 k_{\text{B}}T/l_0^2$) is the spring constant with l_0 being the bond length. The bending rigidity constant is G , σ_i is the diameter of a bead, θ_i is the angle between the monomer bond vectors $(\vec{r}_{i+1} - \vec{r}_i)$ and $(\vec{r}_i - \vec{r}_{i-1})$, and ε is Lennard-Jones energy constant controlling the strength of the excluded volume interaction. The diameter of the SS, $\sigma_{\text{sph}} = 2\sigma_{\text{m}}$, where σ_{m} is the size of the monomer. We chose $N_{\text{m}} = 300$ in our simulations.

Crowders as Spherocylinders. We model the spherocylindrical crowder by connecting five spherical crowders, allowing for overlap (Figure 1a). Five beads in the anisotropic crowders are connected using E_{S} and E_{B} in eq 3 and eq 4, with very large values for the spring and the bending rigidity constants (K_{sp} and G_{sp} , the analogues of K and G in eq 3 and eq 4) in order to maintain the cylindrical shape. We ignore excluded volume interaction between the beads within a particular cylindrical crowder because the parameter G_{sp} is sufficiently large. By choosing the diameter of the cylinder $\sigma_{\text{cyl}} = 2.36\sigma_{\text{m}}$, the volumes of SS and SC crowders are identical ($\pi\sigma_{\text{sph}}^3/6 = \pi\sigma_{\text{cyl}}^3/6 + (\pi\sigma_{\text{cyl}}^2/4) \times 2\sigma_{\text{cyl}}$) (Figure 1a). The parameter values are given in Table S1.

Mixture of SS and SC. To examine if the crowders of different shape have additive effects on the size of the WLC, we also considered a system containing spheres and spherocylinders. We chose an equimolar mixture containing $N_1 = N_2 = N/2$ spheres and spherocylinders so that the total volume fraction of the crowders is $\phi = N_1(v_{\text{SS}} + v_{\text{SC}}) = Nv_{\text{SS}}$ with $v_{\text{SS}} = v_{\text{SC}}$. Thus, in a mixture, $\phi_{\text{SS}} = \phi_{\text{SC}} = \phi/2$, where ϕ_{SS} and ϕ_{SC} are the fractions of volume occupied by SS and SC, respectively.

Modeling the *E. coli* Environment. Using the approximate composition of the crowding particles in *E. coli* in terms of sizes of crowders,⁴² we mimic the cytoplasm as a mixture of spheres containing the three largest particles. They are the ribosome with radius $r_1 = 10.4$ nm, polymerases, and other large complexes with average size $r_2 = 5.2$ nm and smaller complexes where mean size is $r_3 \approx 2.6$ nm. The composition of the three classes of particles is 11, 11, and 8%. These simulations provide a general framework for understanding the fate of stiff molecules in a cell-like environment.

Simulation Details. In order to obtain adequate sampling of the conformational space of the system, we used low friction Langevin dynamics (LFLD). It can be shown, and has been confirmed in

simulations, that the thermodynamic properties of the system do not depend on the choice of the friction coefficient.⁷ In the LFLD, the diameter of the monomer σ_{m} , $\tau = (m\sigma_{\text{m}}^2/\varepsilon)^{1/2}$, and ε is chosen as the units of length, time, and energy, respectively. The value of σ_{m} suitable for DNA is ≈ 3.18 nm. Friction coefficients, ζ_{m} for monomers and ζ_{c} for crowders, were $\zeta_{\text{m}} = 0.05 \text{ m}\tau^{-1}$ and $\zeta_{\text{c}} = \zeta_{\text{m}}\sigma_{\text{sph}}/\sigma_{\text{m}}$ (Table S1).⁵² The duration of each trajectory ranges from 2×10^7 to $5 \times 10^7 \Delta t$, where $\Delta t = 0.01 \tau$.

Initially, a semiflexible chain was placed in a simulation box without the crowders. We performed the LFLD for 10^7 time steps to equilibrate the system. The crowding particles were added to generate a sample with $\phi = 0.05$. Higher volume fractions were reached by inserting additional crowding particles to the simulation box. Subsequently, Lennard-Jones interaction annealing (adiabatic increase of ε) was carried out for $\phi \geq 0.3$ in order to improve the speed of equilibration and to avoid catastrophic crashes during the insertion of particles. To be specific, at the beginning of annealing, we inserted the crowding particles at random positions by assigning $\varepsilon = 0.1 k_{\text{B}}T$ and $0.1 \Delta t$ for the integration time step of LFLD. First, the Lennard-Jones interaction parameter ε was increased by $0.05 k_{\text{B}}T$ for every 10^4 time steps until it reaches $\varepsilon = 1.67 k_{\text{B}}T$. Next, we increased the simulation time step by $0.1 \Delta t$ for every 10^4 time steps until the time step reached Δt .

For purposes of efficient computation, we devised a method (also see Supporting Information in ref 19) in which crowding particles are added on the fly and the volume of the simulation box is adjusted to keep the volume fraction constant. During the simulations, we adjusted the size of the simulation box according to the chain conformation at a given time to minimize the number of crowders. At every time step, we checked if the chain is enclosed in the simulation box. If any monomer and the boundary of the box is closer than <3 times the average distance between the crowders, we resized the box and added crowding particles to the newly extended empty spaces. As a result of constantly resizing the box (a cuboid with changing dimensions), the number of crowders varies. The volume of the cuboid and the number of crowders are varied in such a way that ϕ is a constant. The average number of crowders in our simulations varies from 4000 to 8000 depending on ϕ . In total, we generated 25 trajectories at each volume fraction to obtain statistical properties. We collected data for analysis after a minimum of 10^6 simulation time steps.

The ensemble used in these simulations is not used frequently, although it is discussed by Callen (see page 148 in ref 53). If the system consists of monodisperse particles (generalization to multi-component system follows readily as explicitly shown in ref 53), in which the number of particles (N) and volume (V) fluctuate; the independent variables conjugate to these two variables are chemical potential (μ) and pressure (p). The thermodynamic potential in this ensemble is $\Omega(\mu, p, T) = U - TS + pV - \mu N = G - \mu N = 0$ (follows from Euler relation), which means that variations in N and V do not change the thermodynamic potential $\Omega(\mu, p, T)$. This is precisely what is desired in these simulations.

To ensure that the results do not depend on the choice of the ensemble, we also repeated the simulations in the canonical ensemble for the polydisperse case. As expected on theoretical grounds, the results for R_{g} , $P(R_{\text{g}})$, and the energy per particle (thermodynamic quantity) in the two ensembles are the same. The comparison is given in Figure S6.

■ ASSOCIATED CONTENT

● Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b04531.

Multiple layered neighbor list; shape anisotropy of polymer; analytical expression for the end-to-end distance distribution of WLC; pair correlation function of polydispersed crowders; evidences of equilibration; Figures S1–S5; and Table S1 (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Morelli, M. J.; Allen, R. J.; Rein ten Wolde, P. *Biophys. J.* **2011**, *101*, 2882–2891.
- (2) Tabaka, M.; Kalwarczyk, T.; Holyst, R. *Nucleic Acids Res.* **2014**, *42*, 727–738.
- (3) Ge, X.; Luo, D.; Xu, J. *PLoS One* **2011**, *6*, e28707.
- (4) Zhou, H. X.; Rivas, G.; Minton, A. P. *Annu. Rev. Biophys.* **2008**, *37*, 375.
- (5) Elcock, A. *Curr. Opin. Struct. Biol.* **2010**, *20*, 196–206.
- (6) Cheung, M. S. *Curr. Opin. Struct. Biol.* **2013**, *23*, 212–217.
- (7) Denesyuk, N.; Thirumalai, D. *J. Am. Chem. Soc.* **2011**, *133*, 11858.
- (8) Kilburn, D.; Roh, J. H.; Guo, L.; Briber, R. M.; Woodson, S. A. *J. Am. Chem. Soc.* **2010**, *132*, 8690–8696.
- (9) Pincus, D. L.; Hyeon, C.; Thirumalai, D. *J. Am. Chem. Soc.* **2008**, *130*, 7364–7372.
- (10) Schreiber, G.; Haran, G.; Zhou, H.-X. *Chem. Rev.* **2009**, *109*, 839–860.
- (11) Lerman, L. *Proc. Natl. Acad. Sci. U. S. A.* **1971**, *68*, 1886–1890.
- (12) Dima, R. I.; Thirumalai, D. *J. Phys. Chem. B* **2004**, *108*, 6564–6570.
- (13) Hyeon, C.; Dima, R. I.; Thirumalai, D. *J. Chem. Phys.* **2006**, *125*, 194905.
- (14) Post, C. B.; Zimm, B. H. *Biopolymers* **1979**, *18*, 1487–1501.
- (15) Grosberg, A. Y.; Erukhimovitch, I. Y.; Shakhnovitch, E. *Biopolymers* **1982**, *21*, 2413–2432.
- (16) Castelnovo, M.; Gelbart, W. *Macromolecules* **2004**, *37*, 3510–3517.
- (17) Ramos, J. É. B.; de Vries, R.; Ruggiero Neto, J. J. *Phys. Chem. B* **2005**, *109*, 23661–23665.
- (18) Kudlay, A.; Cheung, M. S.; Thirumalai, D. *J. Phys. Chem. B* **2012**, *116*, 8513–8522.
- (19) Kang, H.; Pincus, P. A.; Hyeon, C.; Thirumalai, D. *Phys. Rev. Lett.* **2015**, *114*, 068303.
- (20) Kim, J.; Jeon, C.; Jeong, H.; Jung, Y.; Ha, B.-Y. *Soft Matter* **2015**, *11*, 1877.
- (21) Shin, J.; Cherstvy, A. G.; Metzler, R. *Soft Matter* **2015**, *11*, 472–488.
- (22) Shendruk, T. N.; Bertrand, M.; de Haan, H. W.; Harden, J. L.; Slater, G. W. *Biophys. J.* **2015**, *108*, 810–820.
- (23) Frisch, H.; Fesciyan, S. J. *Polym. Sci., Polym. Lett. Ed.* **1979**, *17*, 309–315.
- (24) Naghizadeh, J.; Massih, A. R. *Phys. Rev. Lett.* **1978**, *40*, 1299.
- (25) Thirumalai, D. *Phys. Rev. A: At., Mol., Opt. Phys.* **1988**, *37*, 269–276.
- (26) Vasilevskaya, V. V.; Khokhlov, A. R.; Matsuzawa, Y.; Yoshikawa, K. *J. Chem. Phys.* **1995**, *102*, 6595.
- (27) van der Schoot, P. *Macromolecules* **1998**, *31*, 4635–4638.
- (28) Krotova, M. K.; Vasilevskaya, V. V.; Makita, N.; Yoshikawa, K.; Khokhlov, A. R. *Phys. Rev. Lett.* **2010**, *105*, 128302.
- (29) Diamant, H.; Andelman, D. *Phys. Rev. E: Stat. Phys., Plasmas, Fluids, Relat. Interdiscip. Top.* **2000**, *61*, 6740.
- (30) Shaw, M. R.; Thirumalai, D. *Phys. Rev. A: At., Mol., Opt. Phys.* **1991**, *44*, R4797.
- (31) Hyeon, C.; Thirumalai, D. *J. Chem. Phys.* **2006**, *124*, 104905.
- (32) Vafabakhsh, R.; Ha, T. *Science* **2012**, *337*, 1097–1101.
- (33) Le, T. T.; Kim, H. D. *Nucleic Acids Res.* **2014**, *42*, 10786–10794.
- (34) Lieberman-Aiden, E.; et al. *Science* **2009**, *326*, 289.
- (35) Parry, B. R.; Surovtsev, I. V.; Cabeen, M. T.; O'Hern, C. S.; Dufresne, E. R.; Jacobs-Wagner, C. *Cell* **2014**, *156*, 183–194.
- (36) Marko, J. F.; Siggia, E. D. *Macromolecules* **1995**, *28*, 8759–8770.
- (37) Moghaddam, S.; Caliskan, G.; Chauhan, S.; Hyeon, C.; Briber, R.; Thirumalai, D.; Woodson, S. *J. Mol. Biol.* **2009**, *393*, 753–764.
- (38) Asakura, S.; Oosawa, F. *J. Polym. Sci.* **1958**, *33*, 183–192.
- (39) Ogston, A. G. *J. Phys. Chem.* **1970**, *74*, 668–669.
- (40) Vliegthart, G.; Lekkerkerker, H. J. *J. Chem. Phys.* **1999**, *111*, 4153–4157.
- (41) Oversteegen, S.; Wijnhoven, J.; Vonk, C.; Lekkerkerker, H. J. *Phys. Chem. B* **2004**, *108*, 18158–18163.
- (42) Roberts, E.; Magis, A.; Ortiz, J. O.; Baumeister, W.; Luthey-Schulten, Z. *PLoS Comput. Biol.* **2011**, *7*, e1002010.
- (43) Williamson, J. J.; Evans, R. M. L. *Soft Matter* **2013**, *9*, 3600–3612.
- (44) de Gennes, P. G. *Scaling Concepts in Polymer Physics*; Cornell University Press: Ithaca, NY, 1979.
- (45) Brochard, F.; De Gennes, P. *Ferroelectrics* **1980**, *30*, 33–47.
- (46) Xia, Z.; Das, P.; Shakhnovitch, E. I.; Zhou, R. *J. Am. Chem. Soc.* **2012**, *134*, 18266–18274.
- (47) Caliskan, G.; Hyeon, C.; Perez-Salas, U.; Briber, R. M.; Woodson, S. A.; Thirumalai, D. *Phys. Rev. Lett.* **2005**, *95*, 268303.
- (48) Chen, Y.; Abrams, E. S.; Boles, T. C.; Pedersen, J. N.; Flyvbjerg, H.; Austin, R. H.; Sturm, J. C. *Phys. Rev. Lett.* **2015**, *114*, 198303.
- (49) Guzman, I.; Gruebele, M. *J. Phys. Chem. B* **2014**, *118*, 8459–8470.
- (50) Politou, A.; Temussi, P. A. *Curr. Opin. Struct. Biol.* **2015**, *30*, 1–6.
- (51) Frederick, K. B.; Sept, D.; de la Cruz, E. M. *J. Mol. Biol.* **2008**, *378*, 540–550.
- (52) Veitshans, T.; Klimov, D.; Thirumalai, D. *Folding Des.* **1997**, *2*, 1–22.
- (53) Callen, H. B. *Thermodynamics and an Introduction to Thermostatistics*; Wiley: New York, 1985.